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(21) International Application Number: PCT/US98/07584 (22) International Filing Date: 15 April 1998 (15.04.98) (30) Priority Data: 08/842,842 16 April 1997 (16.04.97) US 08/880,855 23 June 1997 (23.06.97) US 09/052,521 30 March 1998 (30.03.98) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 08/880,855 (CIP) Filed on 23 June 1997 (23.06.97) (71) Applicant (for all designated States except US): AMGEN INC. [US/US]; Amgen Center, One Amgen Center Drive, Thousand Oaks, CA 91320-1789 (US). (72) Inventor; and (75) Inventor/Applicant (for US only): BOYLE, William, J. [US/US]; 11678 Chestnut Ridge Street, Moorpark, CA 93021 (US).			(74) Agents: ODRE, Steven, M. et al.; Amgen, Inc., Amgen Center, One Amgen Center Drive, Thousand Oaks, CA 91320-1789 (US). (81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> <i>With an indication in relation to deposited biological material furnished under Rule 13bis separately from the description.</i>
(54) Title: OSTEOPROTEGERIN BINDING PROTEINS AND RECEPTORS			
(57) Abstract <p>A novel polypeptide, osteoprotegerin binding protein, involved in osteoclast maturation has been identified based upon its affinity for osteoprotegerin. Nucleic acid sequences encoding the polypeptide, or a fragment, analog or derivative thereof, vectors and host cells for production, methods of preparing osteoprotegerin binding protein, and binding assays are also described. Compositions and methods for the treatment of bone diseases such as osteoporosis, bone loss due to arthritis or metastasis, hypercalcemia, and Paget's disease are also provided. Receptors for osteoprotegerin binding proteins are also described. The receptors, and agonists and antagonists thereof, may be used to treat bone diseases.</p>			

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OSTEOPROTEGERIN BINDING PROTEINS AND RECEPTORS

Field of the Invention

5 The present invention relates to polypeptides which are involved in osteoclast differentiation. More particularly, the invention relates to osteoprotegerin binding proteins, nucleic acids encoding the proteins, expression vectors and host cells for production of the
10 proteins, and binding assays. Compositions and methods for the treatment of bone diseases, such as osteoporosis, bone loss from arthritis, Paget's disease, and hypercalcemia, are also described.

 The invention also relates to receptors for
15 osteoprotegerin binding proteins and methods and compositions for the treatment of bone diseases using the receptors.

Background of the Invention

20 Living bone tissue exhibits a dynamic equilibrium between deposition and resorption of bone. These processes are mediated primarily by two cell types: osteoblasts, which secrete molecules that comprise the organic matrix of bone; and osteoclasts,
25 which promote dissolution of the bone matrix and solubilization of bone salts. In young individuals with growing bone, the rate of bone deposition exceeds the rate of bone resorption, while in older individuals the rate of resorption can exceed deposition. In the
30 latter situation, the increased breakdown of bone leads to reduced bone mass and strength, increased risk of fractures, and slow or incomplete repair of broken bones.

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Osteoclasts are large phagocytic multinucleated cells which are formed from hematopoietic precursor cells in the bone marrow. Although the growth and formation of mature functional osteoclasts is not well understood, it is thought that osteoclasts mature along the monocyte/macrophage cell lineage in response to exposure to various growth-promoting factors. Early development of bone marrow precursor cells to preosteoclasts are believed to be mediated by soluble factors such as tumor necrosis factor- α (TNF- α), tumor necrosis factor- β (TNF- β), interleukin-1 (IL-1), interleukin-4 (IL-4), interleukin-6 (IL-6), and leukemia inhibitory factor (LIF). In culture, preosteoclasts are formed in the presence of added macrophage colony stimulating factor (M-CSF). These factors act primarily in early steps of osteoclast development. The involvement of polypeptide factors in terminal stages of osteoclast formation has not been extensively reported. It has been reported, however, that parathyroid hormone stimulates the formation and activity of osteoclasts and that calcitonin has the opposite effect, although to a lesser extent.

Recently, a new polypeptide factor, termed osteoprotegerin (OPG), has been described which negatively regulated formation of osteoclasts in vitro and in vivo (see co-owned and co-pending U.S. Serial Nos. 08/577,788 filed December 22, 1995, 08/706,945 filed September 3, 1996, and 08/771,777, filed December 20, 1996, hereby incorporated by reference; and PCT Application No. WO96/26271). OPG dramatically increased the bone density in transgenic mice expressing the OPG polypeptide and reduced the extent of bone loss when administered to ovariectomized rats. An analysis of OPG activity in in vitro osteoclast formation revealed that OPG does not interfere with the growth and differentiation of monocyte/macrophage

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precursors, but more likely blocks the differentiation of osteoclasts from monocyte/macrophage precursors. Thus OPG appears to have specificity in regulating the extent of osteoclast formation.

5 OPG comprises two polypeptide domains having different structural and functional properties. The amino-terminal domain spanning about residues 22-194 of the full-length polypeptide (the N-terminal methionine is designated residue 1) shows homology to other
10 members of the tumor necrosis factor receptor (TNFR) family, especially TNFR-2, through conservation of cysteine rich domains characteristic of TNFR family members. The carboxy terminal domain spanning residues 194-401 has no significant homology to any known
15 sequences. Unlike a number of other TNFR family members, OPG appears to be exclusively a secreted protein and does not appear to be synthesized as a membrane associated form.

Based upon its activity as a negative
20 regulator of osteoclast formation, it is postulated that OPG may bind to a polypeptide factor involved in osteoclast differentiation and thereby block one or more terminal steps leading to formation of a mature osteoclast.

25 It is therefore an object of the invention to identify polypeptides which interact with OPG. Said polypeptides may play a role in osteoclast maturation and may be useful in the treatment of bone diseases.

30 Summary of the Invention

A novel member of the tumor necrosis factor family has been identified from a murine cDNA library expressed in COS cells screened using a recombinant OPG-Fc fusion protein as an affinity probe. The new
35 polypeptide is a transmembrane OPG binding protein which is predicted to be 316 amino acids in length, and

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has an amino terminal cytoplasmic domain, a transmembrane domain, and a carboxy terminal extracellular domain. OPG binding proteins of the invention may be membrane-associated or may be in soluble form.

The invention provides for nucleic acids encoding an OPG binding protein, vectors and host cells expressing the polypeptide, and method for producing recombinant OPG binding protein. Antibodies or fragments thereof which specifically bind OPG binding protein are also provided.

OPG binding proteins may be used in assays to quantitate OPG levels in biological samples, identify cells and tissues that display OPG binding protein, and identify new OPG and OPG binding protein family members. Methods of identifying compounds which interact with OPG binding protein are also provided. Such compounds include nucleic acids, peptides, proteins, carbohydrates, lipids or small molecular weight organic molecules and may act either as agonists or antagonists of OPG binding protein activity.

OPG binding proteins are involved in osteoclast differentiation and the level of osteoclast activity in turn modulates bone resorption. OPG binding protein agonists and antagonists modulate osteoclast formation and bone resorption and may be used to treat bone diseases characterized by changes in bone resorption, such as osteoporosis, hypercalcemia, bone loss due to arthritis metastasis, immobilization or periodontal disease, Paget's disease, osteopetrosis, prosthetic loosening and the like. Pharmaceutical compositions comprising OPG binding proteins and OPG binding protein agonists and antagonists are also encompassed by the invention.

Receptors for OPG binding proteins have also been identified from a marine cDNA library constructed

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from bone marrow cells which bind to a fluorescent-label OPG binding protein. The receptors may be used to identify agonists and antagonists of OPG binding protein interactions with its receptor which may be used to treat bone disease.

Description of the Figures

Figure 1. Structure and sequence of the 32D-F3 insert encoding OPG binding protein. Predicted transmembrane domain and sites for asparagine-linked carbohydrate chains are underlined.

Figure 2. OPG binding protein expression in COS-7 cells transfected with pcDNA/32D-F3. Cells were lipofected with pcDNA/32D-F3 DNA, the assayed for binding to either goat anti-human IgG1 alkaline phosphatase conjugate (secondary alone), human OPG[22-201]-Fc plus secondary (OPG-Fc), or a chimeric ATAR extracellular domain-Fc fusion protein (SATAR-Fc). ATAR is a new member of the TNFR superfamily, and the SATAR-Fc fusion protein serves as a control for both human IgG1 Fc domain binding, and generic TNFR related protein, binding to 32D cell surface molecules.

Figure 3. Expression of OPG binding protein in human tissues. Northern blot analysis of human tissue mRNA (Clontech) using a radiolabeled 32D-F3 derived hybridization probe. Relative molecular mass is indicated at the left in kilobase pairs (kb). Arrowhead on right side indicates the migration of an approximately 2.5 kb transcript detected in lymph node mRNA. A very faint band of the same mass is also detected in fetal liver.

Figure 4. Structure and sequence of the pcDNA/ hu OPGbp 1.1 insert encoding the human OPG

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binding protein. The predicted transmembrane domain and site for asparagine-linked carbohydrate chains are underlined.

5 Figure 5. Stimulation of osteoclast
development in vitro from bone marrow macrophage and
ST2 cell cocultures treated with recombinant murine OPG
binding protein [158-316]. Cultures were treated with
varying concentrations of murine OPG binding protein
10 ranging from 1.6 to 500 ng/ml. After 8-10 days,
cultures were lysed, and TRAP activity was measured by
solution assay. In addition, some cultures were
simultaneously treated with 1, 10, 100, 500, and 1000
ng/ml of recombinant murine OPG [22-401]-Fc protein.
15 Murine OPG binding protein induces a dose-dependent
stimulation in osteoclast formation, whereas OPG [22-
401]-Fc inhibits osteoclast formation.

 Figure 6. Stimulation of osteoclast
20 development from bone marrow precursors in vitro in the
presence of M-CSF and murine OPG binding protein [158-
316]. Mouse bone marrow was harvested, and cultured in
the presence 250, 500, 1000, and 2000 U/ml of M-CSF.
Varying concentrations of OPG binding protein [158-
25 316], ranging from 1.6 to 500 ng/ml, were added to
these same cultures. Osteoclast development was
measured by TRAP solution assay.

 Figure 7. Osteoclasts derived from bone
30 marrow cells in the presence of both M-CSF and OPG
binding protein [158-316] resorb bone in vitro. Bone
marrow cells treated with either M-CSF, OPG binding
protein, or with both factors combined, were plated
onto bone slices in culture wells, and were allowed to
35 develop into mature osteoclasts. The resulting
cultures were then stained with Toluidine Blue (left

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column), or histochemically to detect TRAP enzyme activity (right column). In cultures receiving both factors, mature osteoclasts were formed that were capable of eroding bone as judged by the presence of blue stained pits on the bone surface. This correlated with the presence of multiple large, multinucleated, TRAP positive cells.

Figure 8. Graph showing the whole blood ionized calcium (iCa) levels from mice injected with OPG binding protein, 51 hours after the first injection, and in mice also receiving concurrent OPG administration. OPG binding protein significantly and dose dependently increased iCa levels. OPG (1mg/kg/day) completely blocked the increase in iCa at a dose of OPG binding protein of 5ug/day, and partially blocked the increase at a dose of OPG binding protein of 25ug/day. (*), different to vehicle treated control ($p < 0.05$). (#), OPG treated iCa level significantly different to level in mice receiving that dose of OPG binding protein alone ($p < 0.05$).

Figure 9. Radiographs of the left femur and tibia in mice treated with 0, 5, 25 or 100µg/day of OPG binding protein for 3.5 days. There is a dose dependent decrease in bone density evident most clearly in the proximal tibial metaphysis of these mice, and that is profound at a dose of 100µg/day.

Figure 10. Murine ODAR cDNA sequence and protein sequence. Nucleic acid sequence of the ~2.1 kb cDNA clone is shown, and translation of the 625 residue long open reading frame indicated above. The hydrophobic signal peptide is underlined, and the hydrophobic transmembrane sequence (residues 214-234)

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is in bold. Cysteine residues that comprise the cysteine-rich repeat motifs in the extracellular domain are in bold.

5 Figure 11. Immunofluorescent staining of
ODAR-Fc binding to OPG binding protein transfected
cells. COS-7 cells transfected with OPG binding
protein expression plasmid were incubated with human
IgG Fc (top panel), ODAR-Fc (middle panel) or OPG-Fc
10 (bottom panel). A FITC-labeled goat anti-human IgG Fc
antibody was used as a secondary antibody. Positive
binding cells were examined by confocal microscopy.

 Figure 12 . Effects of ODAR-Fc on the
15 generation of osteoclasts from mouse bone marrow in
vitro. Murine bone marrow cultures were established as
in Example 8 and exposed to OPG binding protein (5
ng/ml) and CSF-1 (30 ng/ml). Various concentrations of
ODAR-Fc ranging from 1500 ng/ml to 65 ng/ml were added.
20 Osteoclast formation was assessed by TRAP cytochemistry
and the TRAP solution assay after 5 days in culture.

 Figure 13. Bone mineral density in mice
after treatment for four days with ODAR-Fc at varying
25 doses. Mice received ODAR-Fc by daily subcutaneous
injection in a phosphate buffered saline vehicle.
Mineral density was determined from bones fixed in 70%
ETOH at the proximal tibial metaphysis mice by
peripheral quantitative computed tomography (pQCT)
30 (XCT-960M, Norland Medical Systems, Ft Atkinson, WI).
Two 0.5 mm cross-sections of bone, 1.5 mm and 2.0 mm
from the proximal end of the tibia were analyzed (XMICE
5.2, Stratec, Germany) to determine total bone mineral
density in the metaphysis. A soft tissue separation
35 threshold of 1500 was used to define the boundary of
the metaphyseal bone. ODAR-Fc produced a significant

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increase in bone mineral density in the proximal tibial metaphysis in a dose dependent manner. Group n = 4.

Detailed Description of the Invention

5 The invention provides for a polypeptide referred to as an OPG binding protein, which specifically binds OPG and is involved in osteoclast differentiation. A cDNA clone encoding the murine form of the polypeptide was identified from a library
10 prepared from a mouse myelomonocytic cell line 32-D and transfected into COS cells. Transfectants were screened for their ability to bind to an OPG[22-201]-Fc fusion polypeptide (Example 1). The nucleic acid sequence revealed that OPG binding protein is a novel
15 member of the TNF family and is most closely related to AGP-1, a polypeptide previously described in co-owned and co-pending U.S. Serial No. 08/660,562, filed June 7, 1996. (A polypeptide identical to AGP-1 and designated TRAIL is described in Wiley et al. Immunity
20 3, 673-682 (1995)). OPG binding protein is predicted to be a type II transmembrane protein having a cytoplasmic domain at the amino terminus, a transmembrane domain, and a carboxy terminal extracellular domain (Figure 1). The amino terminal
25 cytoplasmic domain spans about residues 1-48, the transmembrane domain spans about residues 49-69 and the extracellular domain spans about residues 70-316 as shown in Figure 1 (SEQ ID NO:___). The membrane-associated protein specifically binds OPG (Figure 2).
30 Thus OPG binding protein and OPG share many characteristics of a receptor-ligand pair although it is possible that other naturally-occurring receptors for OPG binding protein exist.

 A DNA clone encoding human OPG binding
35 protein was isolated from a lymph node cDNA library. The human sequence (Figure 4) is homologous to the

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murine sequence. Purified soluble murine OPG binding protein stimulated osteoclast formation in vitro and induced hypercalcemia and bone resorption in vivo.

OPG binding protein refers to a polypeptide
5 having an amino acid sequence of mammalian OPG binding protein, or a fragment, analog, or derivative thereof, and having at least the activity of binding OPG. In preferred embodiments, OPG binding protein is of murine or human origin. In another embodiment, OPG binding
10 protein is a soluble protein having, in one form, an isolated extracellular domain separate from cytoplasmic and transmembrane domains. OPG binding protein is involved in osteoclast differentiation and in the rate and extent of bone resorption, and was found to
15 stimulate osteoclast formation and stimulate bone resorption.

Nucleic Acids

The invention provides for isolated nucleic
20 acids encoding OPG binding proteins. As used herein, the term nucleic acid comprises cDNA, genomic DNA, wholly or partially synthetic DNA, and RNA. The nucleic acids of the invention are selected from the group consisting of:

- 25 a) the nucleic acids as shown in Figure 1 (SEQ ID NO: ____) and Figure 4 (SEQ ID NO: ____);
- b) nucleic acids which hybridize to the polypeptide coding regions of the nucleic acids shown in Figure 1 (SEQ ID NO: ____) and Figure 4 (SEQ ID
30 NO: ____); and remain hybridized to the nucleic acids under high stringency conditions; and
- c) nucleic acids which are degenerate to the nucleic acids of (a) or (b).

Nucleic acid hybridizations typically involve
35 a multi-step process comprising a first hybridization step to form nucleic acid duplexes from single strands

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followed by a second hybridization step carried out under more stringent conditions to selectively retain nucleic acid duplexes having the desired homology. The conditions of the first hybridization step are

5 generally not crucial, provided they are not of higher stringency than the second hybridization step. Generally, the second hybridization is carried out under conditions of high stringency, wherein "high stringency" conditions refers to conditions of

10 temperature and salt which are about 12-20°C below the melting temperature (T_m) of a perfect hybrid of part or all of the complementary strands corresponding to Figure 1 (SEQ. ID. NO: ____) and Figure 4 (SEQ ID NO:____). In one embodiment, "high stringency"

15 conditions refer to conditions of about 65°C and not more than about 1M Na+. It is understood that salt concentration, temperature and/or length of incubation may be varied in either the first or second hybridization steps such that one obtains the

20 hybridizing nucleic acid molecules according to the invention. Conditions for hybridization of nucleic acids and calculations of T_m for nucleic acid hybrids are described in Sambrook et al. Molecular Cloning: A Laboratory Manual Cold Spring Harbor Laboratory Press,

25 New York (1989).

The nucleic acids of the invention may hybridize to part or all of the polypeptide coding regions of OPG binding protein as shown in Figure 1 (SEQ ID NO: ____) and Figure 4 (SEQ ID NO:____); and

30 therefore may be truncations or extensions of the nucleic acid sequences shown therein. Truncated or extended nucleic acids are encompassed by the invention provided that they retain at least the property of binding OPG. In one embodiment, the nucleic acid will

35 encode a polypeptide of at least about 10 amino acids.

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In another embodiment, the nucleic acid will encode a polypeptide of at least about 20 amino acids. In yet another embodiment, the nucleic acid will encode a polypeptide of at least about 50 amino acids. The hybridizing nucleic acids may also include noncoding sequences located 5' and/or 3' to the OPG binding protein coding regions. Noncoding sequences include regulatory regions involved in expression of OPG binding protein, such as promoters, enhancer regions, translational initiation sites, transcription termination sites and the like.

In preferred embodiments, the nucleic acids of the invention encode mouse or human OPG binding protein. Nucleic acids may encode a membrane bound form of OPG binding protein or soluble forms which lack a functional transmembrane region. The predicted transmembrane region for murine OPG binding protein includes amino acid residues 49-69 inclusive as shown in Figure 1 (SEQ. ID. NO: ____). The predicted transmembrane region for human OPG binding protein includes residues 49-69 as shown in Figure 4 (SEQ ID NO: ____). Substitutions which replace hydrophobic amino acid residues in this region with neutral or hydrophilic amino acid residues would be expected to disrupt membrane association and result in soluble OPG binding protein. In addition, deletions of part or all the transmembrane region would also be expected to produce soluble forms of OPG binding protein. Nucleic acids encoding amino acid residues 70-316 as shown in Figure 1 (SEQ ID NO: ____), or fragments and analogs thereof, encompass soluble OPG binding proteins.

Nucleic acids encoding truncated forms of soluble human OPG binding proteins are also included. Soluble forms include residues 69-317 as shown in Figure 4 (SEQ ID NO: ____) and truncations thereof. In one embodiment, N-terminal truncations generate

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polypeptides from residues, 70-317, 71-317, 72-317, and so forth. In another embodiment, nucleic acids encode soluble OPGbp comprising residues 69-317 and N-terminal truncations thereof up to OPGbp [158-317], or
5 alternatively, up to OPGbp [166-317].

Plasmid phuOPGbp 1.1 in E. coli strain DH10 encoding human OPG binding protein was deposited with the American Type Culture Collection, Rockville, MD on June 13, 1997.

10 Nucleic acid sequences of the invention may be used for the detection of sequences encoding OPG binding protein in biological samples. In particular, the sequences may be used to screen cDNA and genomic libraries for related OPG binding protein sequences,
15 especially those from other species. The nucleic acids are also useful for modulating levels of OPG binding protein by anti-sense technology or in vivo gene expression. Development of transgenic animals expressing OPG binding protein is useful for production
20 of the polypeptide and for the study of in vivo biological activity.

Vectors and Host Cells

The nucleic acids of the invention will be
25 linked with DNA sequences so as to express biologically active OPG binding protein. Sequences required for expression are known to those skilled in the art and include promoters and enhancer sequences for initiation of RNA synthesis, transcription termination sites,
30 ribosome binding sites for the initiation of protein synthesis, and leader sequences for secretion. Sequences directing expression and secretion of OPG binding protein may be homologous, i.e., the sequences are identical or similar to those sequences in the
35 genome involved in OPG binding protein expression and secretion, or they may be heterologous. A variety of

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plasmid vectors are available for expressing OPG binding protein in host cells (see, for example, Methods in Enzymology v. 185, Goeddel, D.V. ed., Academic Press (1990)). For expression in mammalian
5 host cells, a preferred embodiment is plasmid pDSRa described in PCT Application No. 90/14363. For expression in bacterial host cells, preferred embodiments include plasmids harboring the lux promoter (see co-owned and co-pending U.S. Serial No.
10 08/577,778, filed December 22, 1995). In addition, vectors are available for the tissue-specific expression of OPG binding protein in transgenic animals. Retroviral and adenovirus-based gene transfer vectors may also be used for the expression of OPG
15 binding protein in human cells for in vivo therapy (see PCT Application No. 86/00922).

Procaryotic and eucaryotic host cells expressing OPG binding protein are also provided by the invention. Host cells include bacterial, yeast, plant,
20 insect or mammalian cells. OPG binding protein may also be produced in transgenic animals such as mice or goats. Plasmids and vectors containing the nucleic acids of the invention are introduced into appropriate host cells using transfection or transformation
25 techniques known to one skilled in the art. Host cells may contain DNA sequences encoding OPG binding protein as shown in Figure 1 or a portion thereof, such as the extracellular domain or the cytoplasmic domain. Nucleic acids encoding OPG binding proteins may be
30 modified by substitution of codons which allow for optimal expression in a given host. At least some of the codons may be so-called preference codons which do not alter the amino acid sequence and are frequently found in genes that are highly expressed. However, it
35 is understood that codon alterations to optimize expression are not restricted to the introduction of

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preference codons. Examples of preferred mammalian host cells for OPG binding protein expression include, but are not limited to COS, CHO⁻, 293 and 3T3 cells. A preferred bacterial host cell is Escherichia coli.

5

Polypeptides

The invention also provides OPG binding protein as the product of procaryotic or eucaryotic expression of an exogenous DNA sequence, i.e., OPG
10 binding protein is recombinant OPG binding protein. Exogenous DNA sequences include cDNA, genomic DNA and synthetic DNA sequences. OPG binding protein may be the product of bacterial, yeast, plant, insect or mammalian cells expression, or from cell-free
15 translation systems. OPG binding protein produced in bacterial cells will have an N-terminal methionine residue. The invention also provides for a process of producing OPG binding protein comprising growing procaryotic or eucaryotic host cells transformed or
20 transfected with nucleic acids encoding OPG binding protein and isolating polypeptide expression products of the nucleic acids.

Polypeptides which are mamalian OPG binding proteins or are fragments, analogs or derivatives
25 thereof are encompassed by the invention. In a preferred embodiment, the OPG binding protein is human OPG binding protein. A fragment of OPG binding protein refers to a polypeptide having a deletion of one or more amino acids such that the resulting polypeptide
30 has at least the property of binding OPG. Said fragments will have deletions originating from the amino terminal end, the carboxy terminal end, and internal regions of the polypeptide. Fragments of OPG binding protein are at least about ten amino acids, at
35 least about 20 amino acids, or at least about 50 amino acids in length. In preferred embodiments, OPG binding

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protein will have a deletion of one or more amino acids from the transmembrane region (amino acid residues 49-69 as shown in Figure 1), or, alternatively, one or more amino acids from the amino-terminus up to and/or including the transmembrane region (amino acid residues 1-49 as shown in Figure 1). In another embodiment, OPG binding protein is a soluble protein comprising, for example, amino acid residues 69-316, or 70-316, or N-terminal or C-terminal truncated forms thereof, which retain OPG binding activity. OPG binding protein is also a human soluble protein as shown in Figure 4 comprising residues 69-317 as shown in Figure 4 and N-terminal truncated forms thereof, e.g., 70-517, 71-517, 71-317, 72-317 and so forth. In a preferred embodiment, the soluble human OPG binding protein comprising residues 69-317 and N-terminal truncation thereof up to OPGbp [158-317], or alternatively up to OPG [166-317].

An analog of an OPG binding protein refers to a polypeptide having a substitution or addition of one or more amino acids such that the resulting polypeptide has at least the property of binding OPG. Said analogs will have substitutions or additions at any place along the polypeptide. Preferred analogs include those of soluble OPG binding proteins. Fragments or analogs may be naturally occurring, such as a polypeptide product of an allelic variant or a mRNA splice variant, or they may be constructed using techniques available to one skilled in the art for manipulating and synthesizing nucleic acids. The polypeptides may or may not have an amino terminal methionine residue

Also included in the invention are derivatives of OPG binding protein which are polypeptides that have undergone post-translational modifications (e.g., addition of N-linked or O-linked carbohydrate chains, processing of N-terminal or

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C-terminal ends), attachment of chemical moieties to the amino acid backbone, chemical modifications of N-linked or O-linked carbohydrate chains, and addition of an N-terminal methionine residue as a result of
5 procaryotic host cell expression. In particular, chemically modified derivatives of OPG binding protein which provide additional advantages such as increased stability, longer circulating time, or decreased immunogenicity are contemplated. Of particular use is
10 modification with water soluble polymers, such as polyethylene glycol and derivatives thereof (see for example U.S. Patent No. 4,179,337). The chemical moieties for derivitization may be selected from water soluble polymers such as polyethylene glycol, ethylene
15 glycol/propylene glycol copolymers, carboxymethylcellulose, dextran, polyvinyl alcohol and the like. The polypeptides may be modified at random positions within the molecule, or at predetermined positions within the molecule and may include one, two,
20 three or more attached chemical moieties. Polypeptides may also be modified at pre-determined positions in the polypeptide, such as at the amino terminus, or at a selected lysine or arginine residue within the polypeptide. Other chemical modifications provided
25 include a detectable label, such as an enzymatic, fluorescent, isotopic or affinity label to allow for detection and isolation of the protein.

OPG binding protein chimeras comprising part or all of an OPG binding protein amino acid sequence
30 fused to a heterologous amino acid sequence are also included. The heterologous sequence may be any sequence which allows the resulting fusion protein to retain the at least the activity of binding OPG. In a preferred embodiment, the carboxy terminal
35 extracellular domain of OPG binding protein is fused to a heterologous sequence. Such sequences include

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heterologous cytoplasmic domains that allow for alternative intracellular signalling events, sequences which promote oligomerization such as the Fc region of IgG, enzyme sequences which provide a label for the polypeptide, and sequences which provide affinity probes, such as an antigen-antibody recognition.

The polypeptides of the invention are isolated and purified from tissues and cell lines which express OPG binding protein, either extracted from lysates or from conditioned growth medium, and from transformed host cells expressing OPG binding protein. OPG binding protein may be obtained from murine myelomonocytic cell line 32-D (ATCC accession no. CRL-11346). Human OPG binding protein, or nucleic acids encoding same, may be isolated from human lymph node or fetal liver tissue. Isolated OPG binding protein is free from association with human proteins and other cell constituents.

A method for the purification of OPG binding protein from natural sources (e.g. tissues and cell lines which normally express OPG binding protein) and from transfected host cells is also encompassed by the invention. The purification process may employ one or more standard protein purification steps in an appropriate order to obtain purified protein. The chromatography steps can include ion exchange, gel filtration, hydrophobic interaction, reverse phase, chromatofocusing, affinity chromatography employing an anti-OPG binding protein antibody or biotin-streptavidin affinity complex and the like.

Antibodies

Antibodies specifically binding the polypeptides of the invention are also encompassed by the invention. The antibodies may be produced by immunization with full-length OPG binding protein,

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soluble forms of OPG binding protein, or a fragment thereof. The antibodies of the invention may be polyclonal or monoclonal, or may be recombinant antibodies, such as chimeric antibodies wherein the murine constant regions on light and heavy chains are replaced by human sequences, or CDR-grafted antibodies wherein only the complementary determining regions are of murine origin. Antibodies of the invention may also be human antibodies prepared, for example, by immunization of transgenic animals capable of producing human antibodies (see, for example, PCT Application No. WO93/12227). The antibodies are useful for detecting OPG binding protein in biological samples, thereby allowing the identification of cells or tissues which produce the protein. In addition, antibodies which bind to OPG binding protein and block interaction with other binding compounds may have therapeutic use in modulating osteoclast differentiation and bone resorption.

Antibodies to the OPG binding protein may be useful in treatment of bone diseases such as, osteoporosis and Paget's disease. Antibodies can be tested for binding to the OPG binding protein in the absence or presence of OPG and examined for their ability to inhibit ligand (OPG binding protein) mediated osteoclastogenesis and/or bone resorption. It is also anticipated that the peptides themselves may act as an antagonist of the ligand:receptor interaction and inhibit ligand-mediated osteoclastogenesis, and peptides of the OPG binding protein will be explored for this purpose as well.

Compositions

The invention also provides for pharmaceutical compositions comprising a therapeutically effective amount of the OPG binding

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protein of the invention together with a pharmaceutically acceptable diluent, carrier, solubilizer, emulsifier, preservative and/or adjuvant. The invention also provides for pharmaceutical compositions comprising a therapeutically effective amount of an OPG binding protein agonist or antagonist. The term "therapeutically effective amount" means an amount which provides a therapeutic effect for a specified condition and route of administration. The composition may be in a liquid or lyophilized form and comprises a diluent (Tris, acetate or phosphate buffers) having various pH values and ionic strengths, solubilizer such as Tween or Polysorbate, carriers such as human serum albumin or gelatin, preservatives such as thimerosal or benzyl alcohol, and antioxidants such as ascorbic acid or sodium metabisulfite. Selection of a particular composition will depend upon a number of factors, including the condition being treated, the route of administration and the pharmacokinetic parameters desired. A more extensive survey of component suitable for pharmaceutical compositions is found in Remington's Pharmaceutical Sciences, 18th ed. A.R. Gennaro, ed. Mack, Easton, PA (1980).

In a preferred embodiment, compositions comprising soluble OPG binding proteins are also provided. Also encompassed are compositions comprising soluble OPG binding protein modified with water soluble polymers to increase solubility, stability, plasma half-life and bioavailability. Compositions may also comprise incorporation of soluble OPG binding protein into liposomes, microemulsions, micelles or vesicles for controlled delivery over an extended period of time. Soluble OPG binding protein may be formulated into microparticles suitable for pulmonary administration.

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Compositions of the invention may be administered by injection, either subcutaneous, intravenous or intramuscular, or by oral, nasal, pulmonary or rectal administration. The route of administration eventually chosen will depend upon a number of factors and may be ascertained by one skilled in the art.

The invention also provides for pharmaceutical compositions comprising a therapeutically effective amount of the nucleic acids of the invention together with a pharmaceutically acceptable adjuvant. Nucleic acid compositions will be suitable for the delivery of part or all of the coding region of OPG binding protein and/or flanking regions to cells and tissues as part of an anti-sense therapy regimen.

Methods of Use

OPG binding proteins may be used in a variety of assays for detecting OPG and characterizing interactions with OPG. In general, the assay comprises incubating OPG binding protein with a biological sample containing OPG under conditions which permit binding to OPG to OPG binding protein, and measuring the extent of binding. OPG may be purified or present in mixtures, such as in body fluids or culture medium. Assays may be developed which are qualitative or quantitative, with the latter being useful for determining the binding parameters (affinity constants and kinetics) of OPG to OPG binding protein and for quantitating levels of biologically active OPG in mixtures. Assays may also be used to evaluate the binding of OPG to fragments, analogs and derivatives of OPG binding protein and to identify new OPG and OPG binding protein family members.

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Binding of OPG to OPG binding protein may be carried out in several formats, including cell-based binding assays, membrane binding assays, solution-phase assays and immunoassays. In general, trace levels of labeled OPG are incubated with OPG binding protein samples for a specified period of time followed by measurement of bound OPG by filtration, electrochemiluminescent (ECL, ORIGEN system by IGEN), cell-based or immunoassays. Homogeneous assay technologies for radioactivity (SPA; Amersham) and time resolved fluorescence (HTRF, Packard) can also be implemented. Binding is detected by labeling OPG or an anti-OPG antibody with radioactive isotopes (^{125}I , ^{35}S , ^3H), fluorescent dyes (fluorescein), lanthanide (Eu^{3+}) chelates or cryptates, orbipyridyl-ruthenium (Ru^{2+}) complexes. It is understood that the choice of a labeled probe will depend upon the detection system used. Alternatively, OPG may be modified with an unlabeled epitope tag (e.g., biotin, peptides, His_6 , myc) and bound to proteins such as streptavidin, anti-peptide or anti-protein antibodies which have a detectable label as described above.

In an alternative method, OPG binding protein may be assayed directly using polyclonal or monoclonal antibodies to OPG binding proteins in an immunoassay. Additional forms of OPG binding proteins containing epitope tags as described above may be used in solution and immunoassays.

Methods for indentifying compounds which interact with OPG binding protein are also encompassed by the invention. The method comprises incubating OPG binding protein with a compound under conditions which permit binding of the compound to OPG binding protein, and measuring the extent of binding. The compound may be substantially purified or present in a crude

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mixture. Binding compounds may be nucleic acids, proteins, peptides, carbohydrates, lipids or small molecular weight organic compounds. The compounds may be further characterized by their ability to increase or decrease OPG binding protein activity in order to determine whether they act as an agonist or an antagonist.

OPG binding proteins are also useful for identification of intracellular proteins which interact with the cytoplasmic domain by a yeast two-hybrid screening process. As an example, hybrid constructs comprising DNA encoding the N-terminal 50 amino acids of an OPG binding protein fused to a yeast GAL4-DNA binding domain may be used as a two-hybrid bait plasmid. Positive clones emerging from the screening may be characterized further to identify interacting proteins. This information may help elucidate a intracellular signaling mechanism associated with OPG binding protein and provide intracellular targets for new drugs that modulate bone resorption.

OPG binding protein may be used to treat conditions characterized by excessive bone density. The most common condition is osteopetrosis in which a genetic defect results in elevated bone mass and is usually fatal in the first few years of life. Osteopetrosis is preferably treated by administration of soluble OPG binding protein.

The invention also encompasses modulators (agonists and antagonists) of OPG binding protein and the methods for obtaining them. An OPG binding protein modulator may either increase or decrease at least one activity associated with OPG binding protein, such as ability to bind OPG or some other interacting molecule or to regulate osteoclast maturation. Typically, an agonist or antagonist may be a co-factor, such as a protein, peptide, carbohydrate, lipid or small

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molecular weight molecule, which interacts with OPG binding protein to regulate its activity. Potential polypeptide antagonists include antibodies which react with either soluble or membrane-associated forms of OPG binding protein, and soluble forms of OPG binding protein which comprise part or all of the extracellular domain of OPG binding protein. Molecules which regulate OPG binding protein expression typically include nucleic acids which are complementary to nucleic acids encoding OPG binding protein and which act as anti-sense regulators of expression.

OPG binding protein is involved in controlling formation of mature osteoclasts, the primary cell type implicated in bone resorption. An increase in the rate of bone resorption (over that of bone formation) can lead to various bone disorders collectively referred to as osteopenias, and include osteoporosis, osteomyelitis, hypercalcemia, osteopenia brought on by surgery or steroid administration, Paget's disease, osteonecrosis, bone loss due to rheumatoid arthritis, periodontal bone loss, immobilization, prosthetic loosening and osteolytic metastasis. Conversely, a decrease in the rate of bone resorption can lead to osteopetrosis, a condition marked by excessive bone density. Agonists and antagonists of OPG binding protein modulate osteoclast formation and may be administered to patients suffering from bone disorders. Agonists and antagonists of OPG binding protein used for the treatment of osteopenias may be administered alone or in combination with a therapeutically effective amount of a bone growth promoting agent including bone morphogenic factors designated BMP-1 to BMP-12, transforming growth factor- β and TGF- β family members, fibroblast growth factors FGF-1 to FGF-10, interleukin-1 inhibitors, TNF α inhibitors, parathyroid hormone, E series

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prostaglandins, bisphosphonates and bone-enhancing minerals such as fluoride and calcium. Antagonists of OPG binding proteins may be particularly useful in the treatment of osteopenia.

5

Receptors for Osteoprotegerin Binding Proteins

The invention also provides for receptors which interact with OPG binding proteins. More particularly, the invention provides for an osteoclast differentiation and activation receptor (ODAR). ODAR is a transmembrane polypeptide which shows highest degree of homology to CD40, a TNF receptor family member. The nucleic acid sequence of murine ODAR and encoded polypeptide is shown in Figure 10. The human homolog of murine ODAR may be readily isolated by hybridization screening of a human cDNA or genomic library with the nucleic acid sequence of Figure 10. Procedures for cloning human ODAR are similar to those described in Example 5 for cloning human OPG binding proteins. The human homolog of the polypeptide shown in Figure 10 has appeared in Anderson et al. (Nature 390, 175-179 (1997)) and is referred to therein as RANK. RANK is characterized as a type I transmembrane protein having homology to TNF receptor family members and is involved in dendritic cell function.

Evidence for the interaction of ODAR and OPG binding protein is shown in Example 13. A soluble form of ODAR (ODAR-Fc fusion protein) prevents osteoclast maturation in vitro (Figure 12) and increases bone density in normal mice after subcutaneous injection (Figure 13). The results are consistent with OPG binding protein interacting with and activating ODAR to promote osteoclast maturation.

Osteoclast development and the rate and extent of bone resorption are regulated by the interaction of OPG binding protein and ODAR. Compounds

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which decrease or block the interaction of OPG binding protein and ODAR are potential antagonists of OPG binding protein activity and may disrupt osteoclast development leading to decreased bone resorption.

5 Alternatively, compounds which increase the interaction of OPG binding protein and ODAR are potential agonists which promote osteoclast development and enhance bone resorption.

A variety of assays may be used to measure
10 the interaction of OPG binding protein and ODAR in vitro using purified proteins. These assays may be used to screen compounds for their ability to increase or decrease the rate or extent of binding to ODAR by OPG binding protein. In one type of assay, ODAR
15 protein can be immobilized by attachment to the bottom of the wells of a microtiter plate. Radiolabeled OPG binding protein (for example, iodinated OPG binding protein) and the test compound(s) can then be added either one at a time (in either order) or
20 simultaneously to the wells. After incubation, the wells can be washed and counted using a scintillation counter for radioactivity to determine the extent of binding to ODAR by OPG binding protein in the presence of the test compound. Typically, the compound will be
25 tested over a range of concentrations, and a series of control wells lacking one or more elements of the test assays can be used for accuracy in evaluation of the results. An alternative to this method involves reversing the "positions" of the proteins, i.e.,
30 immobilizing OPG binding protein to the microtiter plate wells, incubating with the test compound and radiolabeled ODAR, and determining the extent of ODAR binding (see, for example, chapter 18 of *Current Protocols in Molecular Biology*, Ausubel et al., eds.,
35 John Wiley & Sons, New York, NY [1995]).

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As an alternative to radiolabelling, OPG binding protein or ODAR may be conjugated to biotin and the presence of biotinylated protein can then be detected using streptavidin linked to an enzyme, such as horse radish peroxidase [HRP] or alkaline phosphatase [AP], that can be detected colorometrically, or by fluorescent tagging of streptavidin. An antibody directed to OPG binding protein or ODAR that is conjugated to biotin may also be used and can be detected after incubation with enzyme-linked streptavidin linked to AP or HRP

OPG binding protein and ODAR may also be immobilized by attachment to agarose beads, acrylic beads or other types of such inert substrates. The substrate-protein complex can be placed in a solution containing the complementary protein and the test compound; after incubation, the beads can be precipitated by centrifugation, and the amount of binding between OPG binding protein and ODAR can be assessed using the methods described above. Alternatively, the substrate-protein complex can be immobilized in a column and the test molecule and complementary protein passed over the column. Formation of a complex between OPG binding protein and ODAR can then be assessed using any of the techniques set forth above, *i.e.*, radiolabeling, antibody binding, or the like.

Another type of in vitro assay that is useful for identifying a compound which increases or decreases formation of an ODAR/OPG binding protein complex is a surface plasmon resonance detector system such as the Biacore assay system (Pharmacia, Piscataway, NJ). The Biacore system may be carried out using the manufacturer's protocol. This assay essentially involves covalent binding of either OPG binding protein or ODAR to a dextran-coated sensor chip which is

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located in a detector. The test compound and the other complementary protein can then be injected into the chamber containing the sensor chip either simultaneously or sequentially and the amount of complementary protein that binds can be assessed based on the change in molecular mass which is physically associated with the dextran-coated side of the of the sensor chip; the change in molecular mass can be measured by the detector system.

10 In some cases, it may be desirable to evaluate two or more test compounds together for use in increasing or decreasing formation of ODAR/OPG binding protein complex. In these cases, the assays set forth above can be readily modified by adding such additional test compound(s) either simultaneously with, or subsequently to, the first test compound. The remainder of steps in the assay are as set forth above.

15 In vitro assays such as those described above may be used advantageously to screen rapidly large numbers of compounds for effects on complex formation by ODAR and OPG binding protein. The assays may be automated to screen compounds generated in phage display, synthetic peptide and chemical synthesis libraries.

25 Compounds which increase or decrease complex formation of OPG binding protein and ODAR may also be screened in cell culture using ODAR-bearing cells and cell lines. Cells and cell lines may be obtained from any mammal, but preferably will be from human or other primate, canine, or rodent sources. ODAR containing cells such as osteoclasts may be enriched from other cell types by affinity chromatography using publicly available procedures. Attachment of OPG binding protein to ODAR-bearing cells is evaluated in the presence or absence of test compounds and the extent of binding may be determined by, for example, flow

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cytometry using a biotinylated antibody to OPG binding protein. Alternatively, a mouse or human osteoclast culture may be established as described in Example 8 and test compounds may be evaluated for their ability to block osteoclast maturation stimulated by addition of CSF-1 and OPG binding protein. Cell culture assays may be used advantageously to further evaluate compounds that score positive in protein binding assays described above.

10 Compounds which increase or decrease the interaction of OPG binding protein with ODAR may also be evaluated for in vivo activity by administration of the compounds to mice followed by measurements of bone density using bone scanning densitometry or
15 radiography. Procedures for measuring bone density are described in PCT publication WO97/23614 and in Example 13.

 The invention provides for compounds which
20 decrease or block the interaction of OPG binding protein and ODAR and are antagonists of osteoclast formation. Such compounds generally fall into two groups. One group includes those compounds which are derived from OPG binding protein or which interact with
25 OPG binding protein. These have been described above. A second group includes those compounds which are derived from ODAR or which interact with ODAR. Examples of compounds which are antagonists of ODAR include nucleic acids, proteins, peptides,
30 carbohydrates, lipids or small molecular weight organic compounds.

 Antagonists of ODAR may be compounds which bind at or near one or more binding sites for OPG bp in the ODAR extracellular domain and decrease or
35 completely block complex formation. Those regions on ODAR that are involved in complex formation with OPG

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binding protein may be identified by analogy with the structure of the homologous TNF β /TNF-R55 complex which has been described in Banner et al. (Cell 73, 431-445 (1993)). For example, the structure of the TNF β /TNF-R55 complex may be used to identify regions of OPG binding protein and ODAR that are involved in complex formation. Compounds may then be designed which preferentially bind to the regions involved in complex formation and act as antagonists. In one approach set forth in Example 11, peptide antigens were designed for use in raising antibodies to OPG binding protein that act as antagonists. These antibodies are expected to bind to OPG binding protein and block complex formation with ODAR. In a similar approach, peptide antigens based upon ODAR structure may be used to raise anti-ODAR antibodies that act as antagonists.

Anatoginists of ODAR may also bind to ODAR at locations distinct from the binding site(s) for OPG bp and induce conformational changes in ODAR polypeptide that result in decreased or nonproductive complex formation with OPG binding proteins.

In one embodiment, an antagonist is a soluble form of ODAR lacking a functional transmembrane domain. Soluble forms of ODAR may have a deletion of one or more amino acids in the transmembrane domain (amino acids 214-234 as shown in Figure 10). Soluble ODAR polypeptides may have part or all of the extracellular domain and are capable of binding OPG binding protein. Optionally, soluble ODAR may be part of a chimeric protein, wherein part or all of the extracellular domain of ODAR is fused to a heterologous amino acid sequence. In one embodiment, the heterologous amino acid sequence is an Fc region from human IgG.

Modulators (agonists and antagonists) of ODAR may be used to prevent or treat osteopenia, including

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osteoporosis, osteomyelitis, hypercalcemia of malignancy, osteopenia brought on by surgery or steroid administration, Paget's disease, osteonecrosis, bone loss due to rheumatoid arthritis, periodontal bone loss, immobilization, prosthetic loosening and osteolytic metastasis. Agonists and antagonists of ODAR used for the treatment of osteopenias may be administered alone or in combination with a therapeutically effective amount of a bone growth promoting agent including bone morphogenic factors designated BMP-1 to BMP-12, transforming growth factor- β and TGF- β family members, fibroblast growth factors FGF-1 to FGF-10, interleukin-1 inhibitors, TNF α inhibitors, parathyroid hormone, E series prostaglandins, bisphosphonates, estrogens, SERMs and bone-enhancing minerals such as fluoride and calcium. Antagonists of ODAR are particularly useful in the treatment of osteopenia.

The following examples are offered to more fully illustrate the invention, but are not construed as limiting the scope thereof.

Example 1

Identification of a cell line source for an OPG binding protein

Osteoprotegerin (OPG) negatively regulates osteoclastogenesis in vitro and in vivo. Since OPG is a TNFR-related protein, it is likely to interact with a TNF-related family member while mediating its effects. With one exception, all known members of the TNF superfamily are type II transmembrane proteins expressed on the cell surface. To identify a source of an OPG binding protein, recombinant OPG-Fc fusion proteins were used as immunoprobes to screen for OPG

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binding proteins located on the surface of various cell lines and primary hematopoietic cells.

Cell lines that grew as adherent cultures in vitro were treated using the following methods: Cells
5 were plated into 24 well tissue culture plates (Falcon), then allowed to grow to approximately 80% confluency. The growth media was then removed, and the adherent cultures were washed with phosphate buffered saline (PBS) (Gibco) containing 1% fetal calf serum
10 (FCS). Recombinant mouse OPG [22-194]-Fc and human OPG [22-201]-Fc fusion proteins (see U.S. Serial No. 08/706,945 filed September 3, 1996) were individually diluted to 5 ug/ml in PBS containing 1% FCS, then added to the cultures and allowed to incubate for 45 min at
15 0°C. The OPG-Fc fusion protein solution was discarded, and the cells were washed in PBS-FCS solution as described above. The cultures were then exposed to phycoerythrin-conjugated goat F(ab') anti-human IgG secondary antibody (Southern Biotechnology Associates
20 Cat. # 2043-09) diluted into PBS-FCS. After a 30-45 min incubation at 0°C, the solution was discarded, and the cultures were washed as described above. The cells were then analysed by immunofluorescent microscopy to detect cell lines which express a cell surface OPG
25 binding protein.

Suspension cell cultures were analysed in a similar manner with the following modifications: The diluent and wash buffer consisted of calcium- and magnesium-free phosphate buffered saline containing 1%
30 FCS. Cells were harvested from exponentially replicating cultures in growth media, pelleted by centrifugation, then resuspended at 1×10^7 cells/ml in a 96 well microtiter tissue culture plate (Falcon). Cells were sequentially exposed to recombinant OPG-Fc
35 fusion proteins, then secondary antibody as described above, and the cells were washed by centrifugation

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between each step. The cells were then analysed by fluorescence activated cell sorting (FACS) using a Becton Dickinson FACscan.

Using this approach, the murine
5 myelomonocytic cell line 32D (ATCC accession no. CRL-11346) was found to express a surface molecule which could be detected with both the mouse OPG[22-194]-Fc and the human OPG[22-201]-Fc fusion proteins. Secondary antibody alone did not bind to the surface of
10 32D cells nor did purified human IgG1 Fc, indicating that binding of the OPG-Fc fusion proteins was due to the OPG moiety. This binding could be competed in a dose dependent manner by the addition of recombinant murine or human OPG[22-401] protein. Thus the OPG
15 region required for its biological activity is capable of specifically binding to a 32D-derived surface molecule.

Example 2

20 Expression cloning of a murine OPG binding protein

A cDNA library was prepared from 32D mRNA, and ligated into the mammalian expression vector pcDNA3.1(+) (Invitrogen, San Diego, CA). Exponentially
25 growing 32D cells maintained in the presence of recombinant interleukin-3 were harvested, and total cell RNA was purified by acid guanidinium thiocyanate-phenol-chloroform extraction (Chomczynski and Sacchi. Anal. Biochem. 162, 156-159, (1987)). The poly (A+)
30 mRNA fraction was obtained from the total RNA preparation by adsorption to, and elution from, Dynabeads Oligo (dT)25 (Dynal Corp) using the manufacturer's recommended procedures. A directional, oligo-dT primed cDNA library was prepared using the
35 Superscript Plasmid System (Gibco BRL, Gaithersburg, Md) using the manufacturer's recommended procedures.

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The resulting cDNA was digested to completion with Sal I and Not I restriction endonuclease, then fractionated by size exclusion gel chromatography. The highest molecular weight fractions were selected, and then

5 ligated into the polylinker region of the plasmid vector pcDNA3.1(+) (Invitrogen, San Diego, CA). This vector contains the CMV promoter upstream of multiple cloning site, and directs high level expression in eukaryotic cells. The library was then electroporated into

10 competent *E. coli* (ElectroMAX DH10B, Gibco, NY), and titered on LB agar containing 100 ug/ml ampicillin. The library was then arrayed into segregated pools containing approximately 1000 clones/pool, and 1.0 ml cultures of each pool were grown for 16-20 hr at 37°C.

15 Plasmid DNA from each culture was prepared using the Qiagen QiaWell 96 Ultra Plasmid Kit (catalog #16191) following manufacturer's recommended procedures.

Arrayed pools of 32D cDNA expression library were individually lipofected into COS-7 cultures, then

20 assayed for the acquisition of a cell surface OPG binding protein. To do this, COS-7 cells were plated at a density of 1×10^6 per ml in six-well tissue culture plates (Costar), then cultured overnight in DMEM (Gibco) containing 10% FCS. Approximately 2 µg of

25 plasmid DNA from each pool was diluted into 0.5 ml of serum-free DMEM, then sterilized by centrifugation through a 0.2 µm Spin-X column (Costar).

Simultaneously, 10 µl of Lipofectamine (Life Technologies Cat # 18324-012) was added to a separate

30 tube containing 0.5ml of serum-free DMEM. The DNA and Lipofectamine solutions were mixed, and allowed to incubate at RT for 30 min. The COS-7 cell cultures were then washed with serum-free DMEM, and the DNA-lipofectamine complexes were exposed to the cultures

35 for 2-5 hr at 37°C. After this period, the media was

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removed, and replaced with DMEM containing 10%FCS. The cells were then cultured for 48 hr at 37°C.

To detect cultures that express an OPG binding protein, the growth media was removed, and the cells were washed with PBS-FCS solution. A 1.0 ml volume of PBS-FCS containing 5 µg/ml of human OPG[22-201]-Fc fusion protein was added to each well and incubated at RT for 1 hr. The cells were washed three times with PBS-FCS solution, and then fixed in PBS containing 2% paraformaldehyde and 0.2% glutaraldehyde in PBS at RT for 5 min. The cultures were washed once with PBS-FCS, then incubated for 1 hr at 65°C while immersed in PBS-FCS solution. The cultures were allowed to cool, and the PBS-FCS solution was aspirated. The cultures were then incubated with an alkaline-phosphatase conjugated goat anti-human IgG (Fc specific) antibody (SIGMA Product # A-9544) at Rt for 30 min, then washed three-times with 20 mM Tris-Cl (pH 7.6), and 137 mM NaCl. Immune complexes that formed during these steps were detected by assaying for alkaline phosphatase activity using the Fast Red TR/AS-MX Substrate Kit (Pierce, Cat. # 34034) following the manufacturer's recommended procedures.

Using this approach, a total of approximately 300,000 independent 32D cDNA clones were screened, represented by 300 transfected pools of 1000 clones each. A single well was identified that contained cells which acquired the ability to be specifically decorated by the OPG-Fc fusion protein. This pool was subdivided by sequential rounds of sib selection, yeilding a single plasmid clone 32D-F3 (Figure 1). 32D-F3 plasmid DNA was then transfected into COS-7 cells, which were immunostained with either FITC-conjugated goat anti-human IgG secondary antibody alone, human OPG[22-201]-Fc fusion protein plus secondary, or with

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ATAR-Fc fusion protein (ATAR also known as HVEM; Montgomery et al. Cell 87, 427-436 (1996)) (Figure 2). The secondary antibody alone did not bind to COS-7/32D-F3 cells, nor did the ATAR-Fc fusion protein. Only the OPG Fc fusion protein bound to the COS-7/32D-F3 cells, indicating that 32D-F3 encoded an OPG binding protein displayed on the surface of expressing cells.

Example 3

10 OPG Binding Protein Sequence

The 32D-F3 clone isolated above contained an approximately 2.3 kb cDNA insert (Figure 1), which was sequenced in both directions on an Applied Biosystems 15 373A automated DNA sequencer using primer-driven Taq dye-terminator reactions (Applied Biosystems) following the manufacturer's recommended procedures. The resulting nucleotide sequence obtained was compared to the DNA sequence database using the FASTA program (GCG, 20 Univeristy of Wisconsin), and analysed for the presence of long open reading frames (LORF's) using the "Six-way open reading frame" application (Frames) (GCG, Univeristy of Wisconsin). A LORF of 316 amino acid (aa) residues beginning at methionine was detected in 25 the appropriate orientation, and was preceded by a 5' untranslated region of about 150 bp. The 5' untranslated region contained an in-frame stop codon upstream of the predicted start codon. This indicates that the structure of the 32D-F3 plasmid is consistent 30 with its ability to utilize the CMV promotor region to direct expression of a 316 aa gene product in mammalian cells.

The predicted OPG binding protein sequence was then compared to the existing database of known 35 protein sequences using a modified version of the FASTA program (Pearson, Meth. Enzymol. 183, 63-98 (1990)).

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The amino acid sequence was also analysed for the presence of specific motifs conserved in all known members of the tumor necrosis factor (TNF) superfamily using the sequence profile method of (Gribkov et al. Proc. Natl. Acad. Sci. USA 83, 4355-4359 (1987)), as modified by Lüthy et al. Protein Sci. 3, 139-146 (1994)). There appeared to be significant homology throughout the OPG binding protein to several members of the TNF superfamily. The mouse OPG binding protein appear to be most closely related to the mouse and human homologs of both TRAIL and CD40 ligand. Further analysis of the OPG binding protein sequence indicated a strong match to the TNF superfamily, with a highly significant Z score of 19.46.

The OPG binding protein amino acid sequence contains a probable hydrophobic transmembrane domain that begins at a M49 and extends to L69. Based on this configuration relative to the methionine start codon, the OPG binding protein is predicted to be a type II transmembrane protein, with a short N-terminal intracellular domain, and a longer C-terminal extracellular domain (Figure 4). This would be similar to all known TNF family members, with the exception of lymphotoxin alpha (Nagata and Golstein, Science 267, 1449-1456 (1995)).

Example 4

Expression of human OPG binding protein mRNA

Multiple human tissue northern blots (Clontech, Palo Alto, CA) were probed with a ³²P-dCTP labelled 32D-F3 restriction fragment to detect the size of the human transcript and to determine patterns of expression. Northern blots were prehybridized in 5X SSPE, 50% formamide, 5X Denhardt's solution, 0.5% SDS,

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and 100 µg/ml denatured salmon sperm DNA for 2-4 hr at 42°C. The blots were then hybridized in 5X SSPE, 50% formamide, 2X Denhardt's solution, 0.1% SDS, 100 µg/ml denatured salmon sperm DNA, and 5 ng/ml labelled probe
5 for 18-24 hr at 42°C. The blots were then washed in 2X SSC for 10 min at RT, 1X SSC for 10 min at 50°C, then in 0.5X SSC for 10-15 min.

Using a probe derived from the mouse cDNA and hybridization under stringent conditions, a predominant
10 mRNA species with a relative molecular mass of about 2.5 kb was detected in lymph nodes (Figure 3). A faint signal was also detected at the same relative molecular mass in fetal liver mRNA. No OPG binding protein transcripts were detected in the other tissues
15 examined. The data suggest that expression of OPG binding protein mRNA was extremely restricted in human tissues. The data also indicate that the cDNA clone isolated is very close to the size of the native transcript, suggesting 32D-F3 is a full length clone.

20

Example 5

Molecular cloning of the human OPG binding protein

The human homolog of the OPG binding protein
25 is expressed as an approximately 2.5 kb mRNA in human peripheral lymph nodes and is detected by hybridization with a mouse cDNA probe under stringent hybridization conditions. DNA encoding human OPG binding protein is obtained by screening a human lymph node cDNA library
30 by either recombinant bacteriophage plaque, or transformed bacterial colony, hybridization methods (Sambrook et al. Molecular Cloning: A Laboratory Manual Cold Spring Harbor Press, New York (1989)). To this the phage or plasmid cDNA library are screened using
35 radioactively-labeled probes derived from the murine

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OPG binding protein clone 32D-F3. The probes are used to screen nitrocellulose filter lifted from a plated library. These filters are prehybridized and then hybridized using conditions specified in Example 4, ultimately giving rise to purified clones of the human OPG binding protein cDNA. Inserts obtained from any human OPG binding protein clones would be sequenced and analysed as described in Example 3.

A human lymph node poly A⁺ RNA (Clontech, Inc., Palo Alto, CA) was analysed for the presence of OPG-bp transcripts as previously in U.S. Serial No. 08/577,788, filed December 22, 1995. A northern blot of this RNA sample probed under stringent conditions with a 32P-labeled mouse OPG-bp probe indicated the presence of human OPG-bp transcripts. An oligo dT-primed cDNA library was then synthesized from the lymph node mRNA using the SuperScript kit (GIBCO life Technologies, Gaithersburg, MD) as described in example 2. The resulting cDNA was size selected, and the high molecular fraction ligated to plasmid vector pcDNA 3.1 (+) (Invitrogen, San Diego, CA). Electrocompetent E. coli DH10 (GIBCO life Technologies, Gaithersburg, MD) were transformed, and 1 X 10⁶ ampicillin resistant transformants were screened by colony hybridization using a 32P-labeled mouse OPG binding protein probe.

A plasmid clone of putative human OPG binding protein cDNA was isolated, phuOPGbp-1.1, and contained a 2.3 kb insert. The resulting nucleotide sequence of the phuOPGbp-1.1 insert was approximately 80-85% homologous to the mouse OPG binding protein cDNA sequence. Translation of the insert DNA sequence indicated the presence of a long open reading frame predicted to encode a 317 aa polypeptide (Figure 4). Comparison of the mouse and human OPG-bp polypeptides shows that they are ~87% identical, indicating that this protein is highly conserved during evolution.

- 40 -

The human OPG binding protein DNA and protein sequences were not present in Genbank, and there were no homologus EST sequences. As with the murine homolog, the human OPG binding protein shows strong
5 sequence similarity to all members of the TNF α superfamily of cytokines.

Example 6

Cloning and Bacterial Expression of OPG binding protein

10

PCR amplification employing the primer pairs and templates described below are used to generate various forms of murine OPG binding proteins. One primer of each pair introduces a TAA stop codon and a
15 unique XhoI or SacII site following the carboxy terminus of the gene. The other primer of each pair introduces a unique NdeI site, a N-terminal methionine, and optimized codons for the amino terminal portion of the gene. PCR and thermocycling is performed using
20 standard recombinant DNA methodology. The PCR products are purified, restriction digested, and inserted into the unique NdeI and XhoI or SacII sites of vector pAMG21 (ATCC accession no. 98113) and transformed into the prototrophic E. coli 393 or 2596. Other commonly
25 used E. coli expression vectors and host cells are also suitable for expression. After transformation, the clones are selected, plasmid DNA is isolated and the sequence of the OPG binding protein insert is confirmed.

30

pAMG21-Murine OPG binding protein [75-316]

This construct was engineered to be 242 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met(75)-Asp-Pro-Asn-Arg-----
35 Gln-Asp-Ile-Asp(316)-COOH. The template to be used for

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PCR was pcDNA/32D-F3 and oligonucleotides #1581-72 and #1581-76 were the primer pair to be used for PCR and cloning this gene construct.

5 1581-72:

5'-GTTCTCCTCATATGGATCCAAACCGTATTCTGAAGACAGCACTCACTGCTT-3'

(SEQ ID NO:___)

1581-76:

5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3'

10 (SEQ ID NO:___)

pAMG21-Murine OPG binding protein [95-316]

This construct was engineered to be 223 amino acids in length and have the following N-terminal and
15 C-terminal residues, NH₂-Met-His(95)-Glu-Asn-Ala-Gly---
----Gln-Asp-Ile-Asp(316)-COOH. The template used for
PCR was pcDNA/32D-F3 and oligonucleotides #1591-90 and
#1591-95 were the primer pair used for PCR and cloning
this gene construct.

20

1591-90:

5'-ATTGATTCTAGAAGGAGGAATAACATATGCATGAAAACGCAGGTCTGCAG-3'

(SEQ ID NO:___)

1591-95:

25 5'-TATCCGCGGATCCTCGAGTTAGTCTATGTCCTGAACTTTGAA-3'

(SEQ ID NO:___)

pAMG21-Murine OPG binding protein [107-316]

This construct was engineered to be 211 amino
30 acids in length and have the following N-terminal and
C-terminal residues, NH₂-Met-Ser(107)-Glu-Asp-Thr-Leu--
-----Gln-Asp-Ile-Asp(316)-COOH. The template used for
PCR was pcDNA/32D-F3 and oligonucleotides #1591-93 and
#1591-95 were the primer pair used for PCR and cloning
35 this gene construct.

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1591-93:

5'-ATTTGATTCTAGAAGGAGGAATAACATATGTCTGAAGACACTCTGCCGGACTCC-3'

(SEQ ID NO:___)

5 1591-95:

5'-TATCCGCGGATCCTCGAGTTAGTCTATGTCCTGAACTTTGAA-3'

(SEQ ID NO:___)

10 pAMG21-Murine OPG binding protein [118-316]

This construct was engineered to be 199 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met(118)-Lys-Gln-Ala-Phe-Gln-
-----Gln-Asp-Ile-Asp(316)-COOH. The template used for
15 PCR was pcDNA/32D-F3 and oligonucleotides #1591-94 and #1591-95 were the primer pair used for PCR and cloning this gene construct.

1591-94:

20 5'-ATTTGATTCTAGAAGGAGGAATAACATATGAAACAAGCTTTTCAGGGG-3'

(SEQ ID NO:___)

1591-95:

5'-TATCCGCGGATCCTCGAGTTAGTCTATGTCCTGAACTTTGAA-3'

(SEQ ID NO:___)

25

pAMG21-Murine OPG binding protein [128-316]

This construct was engineered to be 190 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met-Lys(128)-Glu-Leu-Gln-His-
-----Gln-Asp-Ile-Asp(316)-COOH. The template used for
30 PCR was pcDNA/32D-F3 and oligonucleotides #1591-91 and #1591-95 were the primer pair used for PCR and cloning this gene construct.

35 1591-91:

5'-ATTTGATTCTAGAAGGAGGAATAACATATGAAAGAACTGCAGCACATTGTG-3'

(SEQ ID NO:___)

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1591-95:

5'-TATCCGCGGATCCTCGAGTTAGTCTATGTCCTGAACTTTGAA-3'

(SEQ ID NO:___)

5 pAMG21-Murine OPG binding protein [137-316]

This construct was engineered to be 181 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met-Gln(137)-Arg-Phe-Ser-Gly-
-----Gln-Asp-Ile-Asp(316)-COOH. The template used for
10 PCR was pcDNA/32D-F3 and oligonucleotides #1591-92 and #1591-95 were the primer pair used for PCR and cloning this gene construct.

1591-92:

15 5'-ATTTGATTCTAGAAGGAGGAATAACATATGCAGCGTTTCTCTGGTGCTCCA-3'

(SEQ ID NO:___)

1591-95:

5'-TATCCGCGGATCCTCGAGTTAGTCTATGTCCTGAACTTTGAA-3'

(SEQ ID NO:___)

20

pAMG21-Murine OPG binding protein [146-316]

This construct is engineered to be 171 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met(146)-Glu-Gly-Ser-Trp-----
25 --Gln-Asp-Ile-Asp(316)-COOH. The template to be used for PCR is pAMG21-murine OPG binding protein [75-316] described above and oligonucleotides #1600-98 and #1581-76 will be the primer pair to be used for PCR and cloning this gene construct.

30

1600-98:

5'- GTTCTCCTCATATGGAAGGTTCTTGTTGGATGTGGCCCA-3'

(SEQ ID NO:___)

1581-76:

35 5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3'

(SEQ ID NO:___)

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pAMG21-Murine OPG binding protein [156-316]

This construct is engineered to be 162 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met-Arg(156)-Gly-Lys-Pro-----
5 --Gln-Asp-Ile-Asp(316)-COOH. The template to be used for PCR is pAMG21-murine OPG binding protein [158-316] below and oligonucleotides #1619-86 and #1581-76 will be the primer pair to be used for PCR and cloning this gene construct.

10

1619-86:

5'-GTTCTCCTCATATGCGTGGTAAACCTGAAGCTCAACCATTTGCA-3'

(SEQ ID NO:___)

1581-76:

15 5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3'

(SEQ ID NO:___)

pAMG21-Murine OPG binding protein [158-316]

This construct was engineered to be 160 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met-Lys(158)-Pro-Glu-Ala-----
20 --Gln-Asp-Ile-Asp(316)-COOH. The template to be used for PCR was pcDNA/32D-F3 and oligonucleotides #1581-73 and #1581-76 were the primer pair to be used for PCR
25 and cloning this gene construct.

1581-73:

5'-GTTCTCCTCATATGAAACCTGAAGCTCAACCATTTGCACACCTCACCATCAAT-3'

(SEQ ID NO:___)

30 1581-76:

5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3'

(SEQ ID NO:___)

pAMG21-Murine OPG binding protein [166-316]

35 This construct is engineered to be 152 amino acids in length and have the following N-terminal and C-terminal residues, NH₂-Met-His(166)-Leu-Thr-Ile-----

- 45 -

--Gln-Asp-Ile-Asp(316)- COOH. The template to be used for PCR is pcDNA/32D-F3 and oligonucleotides #1581-75 and #1581-76 will be the primer pair to be used for PCR and cloning this gene construct.

5

1581-75:

5'-GTTCTCCTCATATGCATTTAACTATTAACGCTGCATCTATCCCAT
CGGGTTCCCATAAAGTCACT-3' (SEQ ID NO:___)

1581-76:

10 5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3' (SEQ ID NO:___)

pAMG21-Murine OPG binding protein [168-316]

This construct is engineered to be 150 amino acids in length and have the following N-terminal and

15 C-terminal residues, NH₂-Met-Thr(168)-Ile-Asn-Ala-----

--Gln-Asp-Ile-Asp(316)- COOH. The template to be used for PCR is pcDNA/32D-F3 and oligonucleotides #1581-74 and #1581-76 will be the primer pair to be used for PCR and cloning.

20

1581-74:

5'-GTTCTCCTCATATGACTATTAACGCTGCATCTATCCCATCGGGTTCCCATAAAGTCACT-3'
(SEQ ID NO:___)

1581-76:

25 5'-TACGCACTCCGCGGTTAGTCTATGTCCTGAACTTTGA-3' (SEQ ID NO:___)

It is understood that the above constructs are examples and one skilled in the art may readily obtain other forms of OPG binding protein using the general methodology presented her.

30

Recombinant bacterial constructs pAMG21-murine OPG binding protein [75-316], [95-316], [107-316], [118-316], [128-316], [137-316], and [158-316] have been cloned, DNA sequence confirmed, and levels of recombinant gene product expression following induction has been examined. All constructs produced levels of recombinant gene product which was readily visible

35

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following SDS polyacrylamide gel electrophoresis and
coomassie staining of crude lysates. Growth of
transformed E. coli 393 or 2596, induction of OPG
binding protein expression and isolation of inclusion
5 bodies containing OPG binding protein is done according
to procedures described in PCT WO97/23614.
Purification of OPG binding proteins from inclusion
bodies requires solubilization and renaturing of OPG
binding protein using procedures available to one
10 skilled in the art. Recombinant murine OPG binding
protein [158-316] was found to be produced mostly
insolubly, but about 40% was found in the soluble
fraction. Recombinant protein was purified from the
soluble fraction as described below and its bioactivity
15 examined.

Example 7

Purification of recombinant murine OPG binding protein [158-316]

20 Frozen bacterial cells harboring expressed murine
OPG binding protein (158-316) were thawed and
resuspended in 20mM tris-HCl pH 7.0, 10mM EDTA. The
cell suspension (20%w/v) was then homogenized by three
25 passes through a microfluidizer. The lysed cell
suspension was centrifuged in a JA14 rotor at 10,000
rpm for 45 minutes. SDS-PAGE analysis showed a band of
approximately 18kd molecular weight present in both
inclusion bodies and the supernatant. The soluble
30 fraction was then applied to a Pharmacia SP Sepharose
4FF column equilibrated with 10mM MES pH 6.0. The OPG
binding protein was eluted with a 20 column volume
gradient of 0-0.4M NaCl in MES pH 6.0. Fractions
containing OPG binding protein were then applied to an
35 ABX Bakerbond column equilibrated with 20mM MES pH 6.0.
OPG binding protein was eluted with a 15CV gradient of

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0-0.5M NaCl in MES pH 6.0. The final product was over 95% homogeneous by SDS-PAGE. N-terminal sequencing gave the following sequence: Met-Lys-Pro-Glu-Ala-Gln-Pro-Phe-Ala-His which was identified to that predicted for a polypeptide starting at residue 158 (with an initiator methionine). The relative molecular weight of the protein during SDS-PAGE does not change upon reduction.

10

Example 8

In vitro bioactivity of recombinant soluble OPG binding protein

Recombinant OPG protein has previously been shown to block vitamin D3-dependent osteoclast formation from bone marrow and spleen precursors in an osteoclast forming assay as described in U.S. Serial No. 08/577,788. Since OPG binding protein binds to OPG, and is a novel member of the TNF family of ligands, it is a potential target of OPG bioactivity. Recombinant soluble OPG binding protein (158-316), representing the minimal core TNF α -like domain, was tested for its ability to modulate osteoclast differentiation from osteoclast precursors. Bone marrow cells were isolated from adult mouse femurs, and treated with M-CSF. The non-adherent fraction was co-cultured with ST2 cells in the presence and absence of both vitamin D3 and dexamethasone. As previously shown, osteoclasts develop only from co-cultures containing stromal cells (ST2), vitamin D3 and dexamethasone. Recombinant soluble OPG binding protein was added at varying concentrations ranging from 0.16 to 500 ng/ml and osteoclast maturation was determined by TRAP solution assay and by visual observation. OPG binding protein strongly stimulated osteoclast

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differentiation and maturation in a dose dependent manner, with half-maximal effects in the 1-2 ng/ml range, suggesting that it acts as an potent inducer of osteoclastogenesis in vitro (Figure 5). The effect of
5 OPG binding protein is blocked by recombinant OPG (Figure 6).

To test whether OPG binding protein could replace the stroma and added steroids, cultures were established using M-CSF at varying concentrations to
10 promote the growth of osteoclast precursors and various amounts of OPG binding protein were also added. As shown in Figure 6, OPG binding protein dose dependently stimulated TRAP activity, and the magnitude of the stimulation was dependent on the level of added M-CSF
15 suggesting that these two factors together are pivotal for osteoclast development. To confirm the biological relevance of this last observation, cultures were established on bovine cortical bone slices and the effects of M-CSF and OPG binding protein either alone
20 or together were tested. As shown in Figure 7, OPG binding protein in the presence of M-CSF stimulated the formation of large TRAP positive osteoclasts that eroded the bone surface resulting in pits. Thus, OPG binding protein acts as an osteoclastogenesis
25 stimulating (differentiation) factor. This suggests that OPG blocks osteoclast development by sequestering OPG binding protein.

Example 9

30 In vivo activity of recombinant
soluble OPG Binding Protein

Based on in vitro studies, recombinant murine OPG binding protein [158-316] produced in E.coli is a
35 potent inducer of osteoclast development from myeloid precursors. To determine its effects in vivo, male

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BDF1 mice aged 4-5 weeks (Charles River Laboratories) received subcutaneous injections of OPG binding protein [158-316] twice a day for three days and on the morning of the fourth day (days 0, 1, 2, and 3). Five groups of mice (n=4) received carrier alone, or 1, 5, 25 or 100µg/ of of OPG binding protein [158-316] per day. An additional 5 groups of mice (n=4) received the above doses of carrier or of OPG binding protein [158-316] and in addition received human Fc-OPG [22-194] at 1mg/Kg/day (approximately 20 µg/day) by single daily subcutaneous injection. Whole blood ionized calcium was determined prior to treatment on day 0 and 3-4 hours after the first daily injection of of OPG binding protein [158-316] on days 1, 2, and 3. Four hours after the last injection on day 3 the mice were sacrificed and radiographs were taken.

Recombinant of OPG binding protein [158-316] produced a significant increase in blood ionized calcium after two days of treatment at dose of 5 µg/day and higher (Figure 8). The severity of the hypercalcemia indicates a potent induction of osteoclast activity resulting from increased bone resorption. Concurrent OPG administration limited hypercalcemia at doses of OPG binding protein [158-316] of 5 and 25 µg/day, but not at 100 µg/day. These same animal were analysed by radiography to determine if there were any effects on bone mineral density visible by X-ray (Figure 9). Recombinant of OPG binding protein [158-316] injected for 3 days decreased bone density in the proximal tibia of mice in a dose-dependent manner. The reduction in bone density was particularly evident in mice receiving 100 µg/d confirming that the profound hypercalcemia in these animals was produced from increased bone resorption and

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the resulting release of calcium from the skeleton. These data clearly indicate that of OPG binding protein [158-316] acts in vivo to promote bone resorption, leading to systemic hypercalcemia, and recombinant OPG
5 abrogates these effects.

Example 10

Cloning and Expression of soluble OPG Binding Protein in mammalian cells

10

The full length clone of murine and human OPG binding protein can be expressed in mammalian cells as previously described in Example 2. Alternatively, the cDNA clones can be modified to encode secreted forms of
15 the protein when expressed in mammalian cells. To do this, the natural 5' end of the cDNA encoding the initiation codon, and extending approximately through the first 69 amino acid of the protein, including the transmembrane spanning region, could be replaced with a
20 signal peptide leader sequence. For example, DNA sequences encoding the initiation codon and signal peptide of a known gene can be spliced to the OPG binding protein cDNA sequence beginning anywhere after the region encoding amino acid residue 68. The
25 resulting recombinant clones are predicted to produce secreted forms of OPB binding protein in mammalian cells, and should undergo post translational modifications which normally occur in the C-terminal extracellular domain of OPG binding protein, such as
30 glycosylation. Using this strategy, a secreted form of OPG binding protein was constructed which has at its 5' end the murine OPG signal peptide, and at its 3' end the human IgG1 Fc domain. The plasmid vector pCEP4/muOPG[22-401]-Fc as described in U.S. Serial No.
35 08/577,788, filed December 22, 1995, was digested with NotI to cleave between the 3' end of OPG and the Fc

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gene. The linearized DNA was then partially digested with XmnI to cleave only between residues 23 and 24 of OPG leaving a blunt end. The restriction digests were then dephosphorylated with CIP and the vector portion
5 of this digest (including residues 1-23 of OPG and Fc) was gel purified.

The murine OPG binding protein cDNA region encoding amino acid residues 69-316 were PCR amplified using Pfu Polymerase (Stratagene, San Diego, CA) from
10 the plasmid template using primers the following oligonucleotides:

1602-61: CCT CTA GGC CTG TAC TTT CGA GCG CAG ATG

1602-59: CCT CTG CGG CCG CGT CTA TGT CCT GAA CTT TG

15 The 1602-61 oligonucleotide amplifies the 5' end of the gene and contains an artificial an StuI site. The 1602-59 primer amplifies the 3' end of the gene and contains an artificial NotI site. The resulting PCR product obtained was digested with NotI
20 and StuI, then gel purified. The purified PCR product was ligated with vector, then used to transform electrocompetent *E. coli* DH10B cells. The resulting clone was sequenced to confirm the integrity of the amplified sequence and restriction site junctions.
25 This plasmid was then used to transfect human 293 fibroblasts, and the OPG binding protein-Fc fusion protein was collected from culture media as previously described in U.S. Serial No. 08/577,788, filed December 22, 1995.

30 Using a similar strategy, an expression vector was designed that is capable of expressing a N-terminal truncation of fused to the human IgG1 Fc domain. This construct consists of the murine OPG signal peptide (aa residue 1-21), fused in frame to
35 murine OPG binding protein residues 158-316, followed by an inframe fusion to human IgG1 Fc domain. To do

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this, the plasmid vector pCEP4/ murine OPG [22-401]
(U.S. Serial No. 08/577,788, filed December 22, 1995),
was digested with HindIII and NotI to remove the entire
OPG reading frame. Murine OPG binding protein,
5 residues 158-316 were PCR amplified using from the
plasmid template pCDNA/32D-F3 using the following
primers:

1616-44: CCT CTC TCG AGT GGA CAA CCC AGA AGC CTG AGG
CCC AGC CAT TTG C
10 1602-59: CCT CTG CGG CCG CGT CTA TGT CCT GAA CTT TG
1616-44 amplifies OPG binding protein
starting at residue 158 as well as containing residues
16-21 of the muOPG signal peptide with an artificial
XhoI site. 1602-59 amplifies the 3' end of the gene
15 and adds an in-frame NotI site. The PCR product was
digested with NotI and XhoI and then gel purified.

The Following complimentary primers were
annealed to eachother to form an adapter encoding the
murine OPG signal peptide and Kozak sequence
20 surrounding the translation initiation site:

1616-41: AGC TTC CAC CAT GAA CAA GTG GCT GTG CTG CGC
ACT CCT GGT GCT CCT GGA CAT CA
25 1616-42: TCG ATG ATG TCC AGG AGC ACC AGG AGT GCG CAG
CAC AGC CAC TTG TTC ATG GTG GA

These primers were annealed, generating 5'
overhangs compatible with HindIII on the 5' end and
30 XhoI on the 3' end. The digested vector obtained
above, the annealed oligos, and the digested PCR
fragment were ligated together and electroporated into
DH10B cells. The resulting clone was sequenced to
confirm authentic reconstruction of the junction
35 between the signal peptide, OPG binding protein
fragment encoding residues 158-316, and the IgG1 Fc

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domain. The recombinant plasmid was purified, transfected into human 293 fibroblasts, and expressed as a conditioned media product as described above.

5 Full length murine and human cDNAs were cloned into the pCEP4 expression vector (Invitrogen, San Diego, CA) then transfected into cultures of human 293 fibroblasts as described in Example 1. The cell cultures were selected with hygromycin as described
10 above and serum-free conditioned media was prepared. The conditioned media was exposed to a column of immobilized recombinant OPG, and shed forms of murine and human recombinant OPG bp were affinity purified. N-terminal sequence analysis of the purified soluble
15 OPG binding proteins indicates that the murine protein is preferentially cleaved before phenylalanine 139, and the human protein is preferentially cleaved before the homologous residue, isoleucine 140. In addition the human protein is also preferentially cleaved before
20 glycine 145. This suggests that naturally occurring soluble forms of human OPG binding protein have amino terminal residues at either isoleucine at position 140 or glycine at position 145.

25 Example 11

Peptides of the OPG binding protein and preparation of polyclonal and monoclonal antibodies to the protein

Antibodies to specific regions of the OPG
30 binding protein may be obtained by immunization with peptides from OPG binding protein. These peptides may be used alone, or conjugated forms of the peptide may be used for immunization.

The crystal structure of mature TNF α has been
35 described [E.Y. Jones, D.I. Stuart, and N.P.C. Walker

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(1990) J. Cell Sci. Suppl. 13, 11-18] and the monomer forms an antiparallel β -pleated sheet sandwich with a jellyroll topology. Ten antiparallel β -strands are observed in this crystal structure and form a beta sandwich with one beta sheet consisting of strands B'BIDG and the other of strands C'CHEF [E.Y. Jones et al., *ibid.*] Two loops of mature TNF α have been implicated from mutagenesis studies to make contacts with receptor, these being the loops formed between beta strand B & B' and the loop between beta strands E & F [C.R. Goh, C-S. Loh, and A.G. Porter (1991) *Protein Engineering* 4, 785-791]. The crystal structure of the complex formed between TNF β and the extracellular domain of the 55kd TNF receptor (TNF-R55) has been solved and the receptor-ligand contacts have been described [D.W. Banner, A. D'Arcy, W. Janes, R. Gentz, H-J. Schoenfeld, C. Broger, H. Loetscher, and W. Lesslauer (1993) *Cell* 73, 431-445]. In agreement with mutagenesis studies described above [C.R. Goh et al., *ibid.*] the corresponding loops BB' and EF of the ligand TNF β were found to make the majority of contacts with the receptor in the resolved crystal structure of the TNF β :TNF-R55 complex. The amino acid sequence of murine OPG binding protein was compared to the amino acid sequences of TNF α and TNF β . The regions of murine OPG binding protein corresponding to the BB' and EF loops were predicted based on this comparison and peptides have been designed and are described below

A. Antigen(s): Recombinant murine OPG binding protein [158-316] has been used as an antigen (ag) for immunization of animals as described below, and serum will be examined using approaches described below. Peptides to the putative BB' and EF loops of

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murine OPG binding protein have been synthesized and will be used for immunization; these peptides are:

BB' loop peptide: NH₂--NAASIPSGSHKVTLSWYHDRGWAKIS--COOH

5 BB' loop-Cys peptide: NH₂--NAASIPSGSHKVTLSWYHDRGWAKISC--
COOH

EF loop peptide: NH₂--VYVVKTSIKIPSSHNLN--COOH

EF loop-Cys peptide: NH₂--VYVVKTSIKIPSSHNLNMC--COOH

10 Peptides with a carboxy-terminal cysteine residue have been used for conjugation using approaches described in section B below, and have been used for immunization.

B. Keyhole Limpet Hemocyanin or Bovine Serum

Albumin Conjugation: Selected peptides or protein

15 fragments may be conjugated to keyhole limpet hemocyanin (KLH) in order to increase their immunogenicity in animals. Also, bovine serum albumin (BSA) conjugated peptides or protein fragments may be utilized in the EIA protocol. Inject Maleimide

20 Activated KLH or BSA (Pierce Chemical Company, Rockford, IL) is reconstituted in dH₂O to a final concentration of 10 mg/ml. Peptide or protein fragments are dissolved in phosphate buffer then mixed with an equivalent mass (g/g) of KLH or BSA. The
25 conjugation is allowed to react for 2 hours at room temperature (rt) with gentle stirring. The solution is then passed over a desalting column or dialyzed against PBS overnight. The peptide conjugate is stored at -
20°C until used in immunizations or in EIAs.

30 C. Immunization: Balb/c mice, (Charles Rivers Laboratories, Wilmington, MA) Lou rats, or New Zealand White rabbits will be subcutaneously injected (SQI) with ag (50 µg, 150 µg, and 100 µg respectively) emulsified in Complete Freund's Adjuvant (CFA, 50%
35 vol/vol; Difco Laboratories, Detroit, MI). Rabbits are then boosted two or three times at 2 week intervals

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with antigen prepared in similar fashion in Incomplete Freund's Adjuvant (ICFA; Difco Laboratories, Detroit, MI). Mice and rats are boosted approximately every 4 weeks. Seven days following the second boost, test
5 bleeds are performed and serum antibody titers determined. When a titer has developed in rabbits, weekly production bleeds of 50 mls are taken for 6 consecutive weeks. Mice and rats are selected for hybridoma production based on serum titer levels;
10 animals with half-maximal titers greater than 5000 are used. Adjustments to this protocol may be applied by one skilled in the art; for example, various types of immunomodulators are now available and may be incorporated into this protocol.

15 D. Enzyme-linked Immunosorbent Assay (EIA): EIAs will be performed to determine serum antibody (ab) titres of individual animals, and later for the screening of potential hybridomas. Flat bottom, high-binding, 96-well microtitration EIA/RIA plates (Costar Corporation, Cambridge, MA) will be coated with
20 purified recombinant protein or protein fragment (antigen, ag) at 5 µg per ml in carbonate-bicarbonate buffer, pH 9.2 (0.015 M Na₂CO₃, 0.035 M NaHCO₃). Protein fragments may be conjugated to bovine serum
25 albumin (BSA) if necessary. Fifty µl of ag will be added to each well. Plates will then be covered with acetate film (ICN Biomedicals, Inc., Costa Mesa, CA) and incubated at room temperature (rt) on a rocking platform for 2 hours or over-night at 4°C. Plates will
30 be blocked for 30 minutes at rt with 250 µl per well 5% BSA solution prepared by mixing 1 part BSA diluent/blocking solution concentrate (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, MD) with 1 part deionized water (dH₂O). Blocking solution having been
35 discarded, 50 µl of serum 2-fold dilutions (1:100 through 1: 12,800) or hybridoma tissue culture

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supernatants will be added to each well. Serum diluent is 1% BSA (10% BSA diluent/blocking solution concentrate diluted 1:10 in Dulbecco's Phosphate Buffered Saline, D-PBS; Gibco BRL, Grand Island, NY)) while hybridoma supernatants are tested undiluted. In the case of hybridoma screening, one well is maintained as a conjugate control, and a second well as a positive ab control. Plates are again incubated at rt, rocking for 1 hour, then washed 4 times using a 1x preparation of wash solution 20x concentrate (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, MD) in dH₂O. Horseradish peroxidase conjugated secondary ab (Boeringer Mannheim Biochemicals, Indianapolis, IN) diluted in 1% BSA is then incubated in each well for 30 minutes. Plates are washed as before, blotted dry, and ABTS peroxidase single component substrate (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, MD) is added. Absorbance is read at 405 nm for each well using a Microplate EL310 reader (Bio-tek Instruments, Inc., Winooski, VT). Half-maximal titre of serum antibody is calculated by plotting the log₁₀ of the serum dilution versus the optical density at 405, then extrapolating at the 50% point of the maximal optical density obtained by that serum. Hybridomas are selected as positive if optical density scores greater than 5-fold above background. Adjustments to this protocol may be applied; in example, conjugated secondary antibody may be chosen for specificity or non-cross-reactivity.

30 E. Cell fusion: The animal selected for hybridoma production is intravenously injected with 50 to 100 µg of ag in PBS. Four days later, the animal is sacrificed by carbon dioxide and its spleen collected under sterile conditions into 35 ml Dulbeccos' Modified Eagle's Medium containing 200 U/ml Penicillin G, 200 µg/ml Streptomycin Sulfate, and 4 mM glutamine (2x

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P/S/G DMEM). The spleen is trimmed of excess fatty tissue, then rinsed through 4 dishes of clean 2x P/S/G DMEM. It is next transferred to a sterile stomacher bag (Tekmar, Cincinnati, OH) containing 10 ml of 2x P/S/G DMEM and disrupted to single cell suspension with the Stomacher Lab Blender 80 (Seward Laboratory UAC House; London, England). As cells are released from the spleen capsule into the media, they are removed from the bag and transferred to a sterile 50 ml conical centrifuge tube (Becton Dickinson and Company, Lincoln Park, NJ). Fresh media is added to the bag and the process is continued until the entire cell content of the spleen is released. These splenocytes are washed 3 times by centrifugation at 225 x g for 10 minutes.

Concurrently, log phase cultures of myeloma cells, Sp2/0-Ag14 or Y3-Ag1.2.3 for mouse or rat splenocyte fusions, respectively, (American Type Culture Collection; Rockville, MD) grown in complete medium (DMEM, 10% inactivated fetal bovine serum, 2 mM glutamine, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, and 10 mM hepes buffer; Gibco Laboratories, Grand Island, NY) are washed in similar fashion. The splenocytes are combined with the myeloma cells and pelleted once again. The media is aspirated from the cell pellet and 2 ml of polyethylene glycol 1500 (PEG 1500; Boehringer Mannheim Biochemicals, Indianapolis, IN) is gently mixed into the cells over the course of 1 minute. Thereafter, an equal volume of 2x P/S/G DMEM is slowly added. The cells are allowed to fuse at 37° C for 2 minutes, then an additional 6 ml of 2x P/S/G DMEM is added. The cells are again set at 37°C for 3 minutes. Finally, 35 ml of 2x P/S/G DMEM is added to the cell suspension, and the cells pelleted by centrifugation. Media is aspirated from the pellet and the cells gently resuspended in complete medium. The cells are distributed over 96-well flat-bottom tissue

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culture plates (Becton Dickinson Labware; Lincoln Park, NJ) by single drops from a 5 ml pipette. Plates are incubated overnight in humidified conditions at 37°C , 5% CO₂. The next day, an equal volume of selection
5 medium is added to each well. Selection consists of 0.1 mM hypoxanthine, 4 x 10⁻⁴ mM aminopterin, and 1.6 x 10⁻² mM thymidine in complete medium. The fusion plates are incubated for 7 days followed by 2 changes of medium during the next 3 days; HAT selection medium is
10 used after each fluid change. Tissue culture supernatants are taken 3 to 4 days after the last fluid change from each hybrid-containing well and tested by EIA for specific antibody reactivity. This protocol has been modified by that in Hudson and Hay, "Practical
15 Immunology, Second Edition", Blackwell Scientific Publications.

Example 12

20 Cloning of an OPG Binding Protein Receptor
Expressed on Hematopoietic Precursor cells

Biologically active recombinant murine OPG binding protein [158-316] was conjugated to fluorescein-isothiocyanate (FITC) to generate a
25 fluorescent probe. Fluorescent labeling was performed by incubation of recombinant murine OPG binding protein [158-316] with 6-fluorescein-5-(and 6) carboxyamido hexanoic acid succinimidyl ester (Molecular Probes, Eugene, OR) at a 1:6 molar ratio for 12 hrs. at 4°C.
30 FITC-labeled OPG binding protein [158-316] was further purified by gel filtration chromatography. Mouse bone marrow cells were isolated and incubated in culture in the presence of CSF-1 and OPG binding protein [158-316] as described in Example 10. Mouse bone marrow cells
35 were cultured overnight in CSF-1 (30 ng/ml) and OPG binding protein [158-316] (20 ng/ml). Non-adherent

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cells were removed first and stored on ice and the remaining adherent cells were removed by incubating with cell dissociation buffer (Sigma Chemicals, St. Louis, MO), pooled with the non-adherent population, and then stained with FITC-OPG binding protein as described above. After washing and resuspending in PBS with 0.5% BSA, the cells were exposed to FITC-OPG binding protein, washed, then sorted by FACS. The population of cells that were positive for staining with the FITC-OPG binding protein was collected and mRNA was isolated as described in Example 2. This mRNA preparation was used to make a cDNA library following procedures described in Example 2.

The cDNA library produced from this source was used for random EST sequence analysis as previously described in PCT Publication No. WO97/23614 and in Simonet et al. (Cell 89, 309-319 (1997)). Using this method, an ~2.1 kb cDNA was detected that encoded a novel TNFR-related protein. The long open reading frame of the murine ODAR cDNA encodes a protein of 625 amino acid residues and contains the hallmark features of TNFR-related proteins: a hydrophobic signal peptide at its N-termini, four tandem cysteine-rich repeat sequences, a hydrophobic transmembrane domain, and a cytoplasmic signalling domain. The homology of this protein with other members of the TNF receptor family and its expression in bone marrow cells that bind FITC-labelled OPG binding protein suggest that it is a potential receptor for the TNF-related OPG binding protein. This protein is designated ODAR, or osteoclast differentiation and activation receptor. The nucleic acid sequence and predicted amino acid sequence of murine ODAR is shown in Figure 10.

Recent analysis of sequences in publicly available databases indicate that this protein is the

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murine homolog of a human TNFR-related protein known as RANK (Anderson et al., Nature 390, 175-179 (1997)).

Example 13

5 Production of Recombinant ODAR Protein in Mammalian Cells

A soluble ODAR extracellular domain fused to the Fc region of human IgG₁ was produced using
10 procedures for the construction and expression of Fc fusion proteins as previously described in WO97/23614 and in Simonet et al., supra. To generate soluble ODAR protein in mammalian cells, cDNA encoding extracellular domain of murine ODAR (amino acids 27-211) was PCR
15 amplified with the following set of oligonucleotide primers:

5' TCT CCA AGC TTG TGA CTC TCC AGG TCA CTC C-3'

(SEQ ID NO:___)

20 5' TCT CCG CGG CCG CGT AAG CCT GGG CCT CAT TGG GTG-3'

(SEQ ID NO:___)

PCR reactions were carried in a volume of 50 µl with 1 unit of vent DNA polymerase (New England
25 Biolabs) in 20 mM Tris-HCl pH 8.8, 10 mM KCl, 10 mM (NH₄)₂SO₄, 0.1% Triton-X100, 10 µM of each dNTP, 1µM of each primer and 10 ng of ODAR cDNA template. Reactions were performed in 94°C for 30 s, 55°C for 30 s, and 72°C for 1 min, for a total of 16 cycles. The PCR fragment
30 was isolated by electrophoresis. The PCR fragment creates a Hind III restriction site at 5' end and a Not I restriction site at 3' end. The Hind III-Not I digested PCR fragment was then subcloned in-frame into a modified pCEP4-Fc vector in front of the human IgG-γ1
35 heavy chain sequence as described previously in

- 62 -

WO97/23614 and in Simonet et al. supra). A linker was introduced which encodes two irrelevant amino acids spanning the junction between the ODAR extracellular domain and the IgG Fc region.

5 The construct was then digested with Nhe I and Hind III and the following annealed oligonucleotide pair encoding OPG signal peptide (amino acid 1-21) was inserted in-frame:

10 5'CTA GCA CCA TGA ACA AGT GGC TGT GCT GCG CAC TCC TGG
TGC TCC TGG ACA TCA TTG AAT GGA CAA CCC AGA-3' (SEQ ID
NO:___)
5'AGC TTC TGG GTT GTC CAT TCA ATG ATG TCC AGG AGC ACC
AGG AGT GCG CAG CAC AGC CAC TTG TTC ATG GTG-3' (SEQ ID
15 NO:___)

A linker which encodes two irrelevant amino acids was introduced between OPG signal peptide and ODAR sequences. The final engineered construct (ODAR-

20 Fc/pCEP4) encodes a fusion protein containing from amino terminus to carboxy terminus: OPG signal peptide (amino acids 1-21)-linker (LysLeu)-ODAR (amino acids 27-211)-linker (AlaAla)-human IgG Fc.

25 The construct was transfected into 293-EBNA-1 cells by calcium phosphate method as described (Ausubel et al., Curr. Prot. Mol. Biol. 1, 9.1.1-9.1.3, (1994). The transfected cells were then selected in 200 µg/ml hygromycin (GibcoBRL) and the resulting drug-resistant
30 mass cultures were pooled and grown to confluence. The cells were washed in PBS once and then cultured in serum-free media for 72 hr. The conditioned media was collected. The ODAR-Fc fusion protein in the media was detected by western blot analysis with anti-human IgG
35 Fc antibody.

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The Fc fusion protein was purified by protein-A column chromatography (Pierce) using the manufacturer's recommended procedures. Fifty pmoles of the purified protein was then subjected to N-terminal
5 sequence analysis by automated Edman degradation as essentially described by Matsudaira et al (J. Biol. Chem. 262, 10-35 (1987)). The following amino acid sequence was read after 10 cycles:

NH₂- K L V T L Q V T P-CO₂H.

10

The binding activity of ODAR-Fc with OPG binding protein was examined by immunofluorescent staining of transfected COS-7 cell cultures as described in Example 2. COS-7 cells were lipofected
15 with 1µg of an expression vector containing DNA encoding murine OPG binding protein. After 48 hr incubation, cells were then incubated in PBS-FBS solution containing 10 mg/µl of human IgG Fc, ODAR-Fc, or OPG-Fc protein at 4°C for 1 hr. Cells were then
20 washed with PBS twice and then incubated in PBS-FBS solution containing 20 µg/ml FITC-labeled goat anti-human IgG (Southern Biotech Associates) for another hr. After washing with PBS, cells were examined by confocal microscopy (ACAS, Ultima, Insight Biomedical Imaging,
25 Inc., Okemos, MI). Both ODAR-Fc and OPG-Fc bind to OPGL transfected COS-7 cells (Figure 11).

Example 14

In vitro biological activity of recombinant soluble

30

ODAR

The ability of ODAR to inhibit stimulation of osteoclast formation by OPG binding protein was assessed in a mouse bone marrow culture in the presence
35 of CSF-1 (30ng/ml) and OPG binding protein (5ng/ml).

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Procedures for the use of mouse bone marrow cultures to study osteoclast maturation are described in WO97/23614 and in Example 8. ODAR-Fc fusion protein produced as described in Example 12 was added to concentrations of 5 65 to 1500ng/ml. Osteoclast formation was assessed by tartrate resistant alkaline phosphatase (TRAP) cytochemistry and the TRAP solution assay after five days in culture.

A dose dependent inhibition of osteoclast 10 formation by ODAR-Fc fusion was observed both by cytochemistry and by TRAP activity (Figure 12). ODAR-Fc fusion protein inhibited osteoclast formation with an ED₅₀ of about 10-50ng/ml.

15 Example 15

In vivo biological activity of recombinant soluble ODAR

Young rapidly growing male BDF1 mice aged 3-4 weeks received varying doses of ODAR-Fc fusion protein 20 by single daily subcutaneous injection in carrier (PBS/0.1% BSA) for four days. The mice were x-rayed on day 5. Doses of ODAR-Fc fusion protein used were 0.5, 1.5 and 5mg/kg/day. For each treatment, all the mice in that group and in the control group that received 25 PBS/0.1% BSA were x-rayed on a single film. The proximal tibial metaphyseal region was compared between pairs of control and treated tibias and scored as a "+" if the treated tibia was denser by visual assessment than the control giving the 8 scores shown below. An 30 arbitrary score of 5/8 was required for a "positive" result. (Dose is in mg/Kg/day). (n=4).

After sacrifice the the right tibia was removed from each animal and the bone density in the proximal tibial metaphysis was measured by peripheral 35 quantitative computerized tomography (pQCT) (Stratec, Germany). Two 0.5 mm cross-sections of bone, 1.5 mm and

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2.0 mm from the proximal end of the tibia were analyzed (XMICE 5.2, Stratec, Germany) to determine total bone mineral density in the metaphysis. A soft tissue separation threshold of 1500 was used to define the boundary of the metaphyseal bone.

ODAR-Fc administration in young growing mice inhibited bone resorption at the proximal tibial growth plate producing a region of increased bone density that was evident visually on radiographs. Radiographic changes were apparent at a dose of 1.5mg/kg/day and above in two experiments (Table 1). Measurement of the bone density by pQCT in samples from the second experiment in a similar region of the tibia confirmed the dose dependent increased in bone density in these mice (Figure 13).

Table 1
Inhibition of bone resorption by ODAR-Fc fusion protein

Experiment #1

Factor	Dose	1	2	3	4	5	6	7	8	Result
ODAR-Fc	5.0	+	+	+	+	+	+	+	+	Positive 8/8
ODAR-Fc	1.5	-	+	+	-	+	+	+	+	Positive 6/8
ODAR-Fc	0.5	-	-	-	-	-	-	-	-	Negative 0/8
ODAR-Fc	0.15	-	-	-	-	-	-	-	-	Negative 0/8

Experiment #2

Factor	Dose	1	2	3	4	5	6	7	8	Result
ODAR-Fc	5.0	+	+	+	+	+	+	+	+	Positive 8/8
ODAR-Fc	1.5	+	+	+	+	+	+	+	+	Positive 8/8
ODAR-Fc	0.5	-	-	-	+	-	-	-	-	Negative 1/8

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* * *

While the present invention has been
5 described in terms of the preferred embodiments, it is
understood that variations and modifications will occur
to those skilled in the art. Therefore, it is intended
that the appended claims cover all such equivalent
variations which come within the scope of the invention
10 as claimed.

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WHAT IS CLAIMED IS:

1. An isolated nucleic acid encoding an osteoprotegerin binding protein selected from the group consisting of:
 - a) the nucleic acid sequence as in Figure 1 (SEQ ID NO:___) and Figure 4 (SEQ ID NO:___);
 - b) nucleic acids which hybridize to the polypeptide coding regions as shown in Figure 1 (SEQ ID NO:___) and Figure 4 (SEQ ID NO:___) and remain hybridized under high stringency conditions; and
 - c) nucleic acids which are degenerate to the nucleic acids of (a) or (b).
2. The nucleic acid of Claim 1 which is cDNA, genomic DNA, synthetic DNA or RNA.
3. A polypeptide encoded by the nucleic acid of Claim 1.
4. The nucleic acid of Claim 1 including one or more codons preferred for Escherichia coli expression.
5. The nucleic acid of Claim 1 having a detectable label attached thereto.
6. A nucleic acid encoding a polypeptide comprising the amino acid sequence of residues 1-316 and residues 70-316 as shown in Figure 1 (SEQ ID NO:___).
7. A nucleic acid encoding a polypeptide comprising amino acid sequence of residues 1-317 and residues 69-317 as shown in Figure 4 (SEQ ID NO:___);

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8. A nucleic acid encoding a soluble osteoprotegerin binding protein.

5 9. The nucleic acid of Claim 8 encoding a polypeptide comprising residues 69-317 as shown in Figure 4 (SEQ ID NO:____) and truncations thereof;

10 10. An expression vector comprising the nucleic acid of Claims 1 and 9.

15 11. The expression vector of Claim 10 wherein the nucleic acid comprises the polypeptide-encoding region as shown in Figure 1 (SEQ ID NO:____) and Figure 4 (SEQ ID NO:____);

12. A host cell transformed or transfected with the expression vector of Claim 10.

20 13. The host cell of Claim 12 which is a eucaryotic or procaryotic cell.

25 14. The host cell of Claim 13 which is Escherichia coli.

15. A process for the production of an osteoprotegerin binding protein comprising:
 growing under suitable nutrient conditions host cells transformed or transfected with
30 the nucleic acid of Claim 1; and
 isolating the polypeptide product of the expression of the nucleic acid.

35 16. A polypeptide produced by the process of Claim 15.

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17. A purified and isolated osteoprotegerin binding protein, or fragment, analog, or derivative thereof.

5

18. The protein of Claim 17 which is a human osteoprotegerin.

19. The protein of Claim 17 having the amino acid sequence as shown in Figure 1 (SEQ ID NO:___) and Figure 4 (SEQ ID NO:___).

10

20. The protein of Claim 17 which has been covalently modified with a water-soluble polymer.

15

21. The protein of Claim 20 wherein the polymer is polyethylene glycol.

22. The protein of Claim 17 which is a soluble osteoprotegerin binding protein.

20

23. The protein of Claim 22 comprising the amino acid sequence from residues 70-316 inclusive as shown in Figure 1 (SEQ ID NO: ___), or a fragment, analog, or derivative thereof.

25

24. The protein of Claim 22 comprising the amino acid sequence from residues 69-317 inclusive as shown in Figure 4 (SEQ ID NO:___) and truncations thereof.

30

25. An antibody or fragment thereof which specifically binds an osteoprotegerin binding protein.

26. The antibody of Claim 25 which is a monoclonal antibody.

35

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27. A method for detecting the presence of an osteoprotegerin binding protein in a biological sample comprising:

- 5 incubating the sample with the antibody of Claim 25 under conditions that allow binding of the antibody to the osteoprotegerin binding protein; and
 detecting the bound antibody.

- 10 28. A method for detecting the presence of osteoprotegerin in a biological sample comprising:
 incubating the sample with an osteoprotegerin binding protein under conditions that allow binding of the protein to osteoprotegerin; and
15 measuring the bound osteoprotegerin binding protein.

29. A method to assess the ability of a candidate compound to bind to an osteoprotegerin
20 binding protein comprising:
 incubating the osteoprotegerin binding protein with the candidate compound under conditions that allow binding; and
 measuring the bound compound.

- 25 30. The method of Claim 29 wherein the compound is an agonist or an antagonist of an osteoprotegerin binding protein.

- 30 31. A method of regulating expression of an osteoprotegerin binding protein in an animal comprising administering to the animal a nucleic acid complementary to the nucleic acids as shown in Figure 1 (SEQ ID NO:___) and Figure 4 (SEQ ID NO: ____).

35

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32. A pharmaceutical composition comprising a therapeutically effective amount of an osteoprotegerin binding protein in a pharmaceutically acceptable carrier, adjuvant, solubilizer, stabilizer and/or
5 anti-oxidant.

33. The composition of Claim 32 wherein the osteoprotegerin binding protein is a human osteoprotegerin binding protein.
10

34. A method of preventing or treating bone disease in a mammal comprising administering a therapeutically effective amount of a modulator of an osteoprotegerin binding protein.
15

35. The method of Claim 34 wherein the modulator is a soluble form of an osteoprotegerin binding protein.

20 36. The method of Claim 35 wherein the modulator is an antibody, or fragment thereof, which specifically binds an osteoprotegerin binding protein.

37. The protein of Claim 22 comprising the
25 amino acid sequence from residues 140-316 inclusive as shown in Figure 4 (SEQ ID NO. __) or a fragment, analog or derivative thereof.

38. The protein of Claim 22 comprising the
30 amino acid sequence from residues 145-316 inclusive as shown in Figure 4 (SEQ ID NO. __) or a fragment, analog or derivative thereof.

39. A method of preventing or treating bone
35 disease in a mammal comprising administering a

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therapeutically effective amount of a modulator of an osteoclast differentiation and activation receptor.

40. The method of Claim 39 wherein the
5 modulator is a soluble form of an osteoclast differentiation and activation receptor.

41. The method of Claim 39 wherein the
modulator is an antibody, or fragment thereof, which
10 specifically binds an osteoclast differentiation and activation factor.

42. A method to assess the ability of a test
compound to increase or decrease binding of
15 osteoprotegerin binding protein to ODAR comprising:
incubating osteoprotegerin binding protein,
ODAR and optionally the test compound under conditions
that allow binding of osteoprotegerin binding protein
to ODAR; and
20 measuring the binding of osteoprotegerin
binding protein to ODAR in the absence and presence of
the test compound.

25

1 / 3 0

GAGCTCGGAT CCACTACTCG ACCCAGCGGT CCGGCCAGGA CCTCTGTGAA CCGGTCGGGG 60
 CCGGGGCCGC CTGGCCGGGA GTCTGCTCGG CCGTGGGTGG CCGAGGAAGG GAGAGAACGA 120
 TCGCGGAGCA GGGCGCCCGA ACTCCGGGCG CCGCGCC ATG CGC CGG GCC AGC CGA 175
 Met Arg Arg Ala Ser Arg
 1 5
 GAC TAC GGC AAG TAC CTG CGC AGC TCG GAG GAG ATG GGC AGC GGC CCC 223
 Asp Tyr Gly Lys Tyr Leu Arg Ser Ser Glu Glu Met Gly Ser Gly Pro
 10 15 20
 GGC GTC CCA CAC GAG GGT CCG CTG CAC CCC GCG CCT TCT GCA CCG GCT 271
 Gly Val Pro His Glu Gly Pro Leu His Pro Ala Pro Ser Ala Pro Ala
 25 30 35
 CCG GCG CCG CCA CCC GCC GCC TCC CGC TCC ATG TTC CTG GCC CTC CTG 319
 Pro Ala Pro Pro Ala Ala Ser Arg Ser Met Phe Leu Ala Leu Leu
 40 45 50

FIG.1A

2 / 3 0

GGG CTG GGA CTG GGC CAG GTG GTC TGC AGC ATC GCT CTG TTC CTG TAC	367
Gly Leu Gly Leu Gly Gln Val Val Cys Ser Ile Ala Leu Phe Leu Tyr	
55 60 65 70	
TTT CGA GCG CAG ATG GAT CCT AAC AGA ATA TCA GAA GAC AGC ACT CAC	415
Phe Arg Ala Gln Met Asp Pro Asn Arg Ile Ser Glu Asp Ser Thr His	
75 80 85	
TGC TTT TAT AGA ATC CTG AGA CTC CAT GAA AAC GCA GGT TTG CAG GAC	463
Cys Phe Tyr Arg Ile Leu Arg Leu His Glu Asn Ala Gly Leu Gln Asp	
90 95 100	
TCG ACT CTG GAG AGT GAA GAC ACA CTA CCT GAC TCC TGC AGG AGG ATG	511
Ser Thr Leu Glu Ser Glu Asp Thr Leu Pro Asp Ser Cys Arg Arg Met	
105 110 115	
AAA CAA GCC TTT CAG GCG GCC GTG CAG AAG GAA GAA CTG CAA CAC ATT GTG	559
Lys Gln Ala Phe Gln Gly Ala Val Gln Lys Glu Leu Gln His Ile Val	
120 125 130	

FIG.1B

3 / 3 0

GGG CCA CAG CGC TTC TCA GGA GCT CCA GCT ATG ATG GAA GGC TCA TGG	607
Gly Pro Gln Arg Phe Ser Gly Ala Pro Ala Met Met Glu Gly Ser Trp	150
135	140
	145
TTG GAT GTG GCC CAG CAG CGA GGC AAG CCT GAG GCC CAG CCA TTT GCA CAC	655
Leu Asp Val Ala Gln Arg Gly Lys Pro Gln Ala Gln Pro Phe Ala His	165
155	160
CTC ACC ATC AAT GCT GCC AGC ATC CCA TCG GGT TCC CAT AAA GTC ACT	703
Leu Thr Ile Asn Ala Ala Ser Ile Pro Ser Gly Ser His Lys Val Thr	180
170	175
CTG TCC TCT TGG TAC CAC GAT CGA GGC TGG GCC AAG ATC TCT AAC ATG	751
Leu Ser Ser Trp Trp Tyr His Asp Arg Gly Trp Ala Lys Ile Ser <u>Asn</u> Met	195
185	190

FIG.1C

4 / 3 0

ACG TTA AGC AAC GGA AAA CTA AGG GTT AAC CAA GAT GGC TTC TAT TAC	799
Thr Leu Ser Asn Gly Lys Leu Arg Val Asn Gln Asp Gly Phe Tyr Tyr	
200 205 210	
CTG TAC GCC AAC ATT TGC TTT CGG CAT CAT GAA ACA TCG GGA AGC GTA	847
Leu Tyr Ala Asn Ile Cys Phe Arg His His Glu Thr Ser Gly Ser Val	
215 220 225 230	
CCT ACA GAC TAT CTT CAG CTG ATG GTG TAT GTC GTT AAA ACC AGC ATC	895
Pro Thr Asp Tyr Leu Gln Leu Met Val Tyr Val Val Lys Thr Ser Ile	
235 240 245	
AAA ATC CCA AGT TCT CAT AAC CTG ATG AAA GGA GGG AGC ACG AAA AAC	943
Lys Ile Pro Ser Ser His Asn Leu Met Lys Gly Gly Ser Thr Lys <u>Asn</u>	
250 255 260	
TGG TCG GGC AAT TCT GAA TTC CAC TTT TAT TCC ATA AAT GTT GGG GGA	991
Trp Ser Gly Asn Ser Glu Phe His Phe Tyr Ser Ile Asn Val Gly Gly	
265 270 275	

FIG.1D

TTT TTC AAG CTC CGA GCT GGT GAA GAA ATT AGC ATT CAG GTG TCC AAC 1039
 Phe Phe Lys Leu Arg Ala Gly Glu Glu Ile Ser Ile Gln Val Ser Asn
 280 285 290
 CCT TCC CTG CTG GAT CCG GAT CAA GAT GCG ACG TAC TTT GGG GCT TTC 1087
 Pro Ser Leu Leu Asp Pro Asp Gln Asp Ala Thr Tyr Phe Gly Ala Phe
 295 300 305 310
 AAA GTT CAG GAC ATA GAC T GAGACTCATT TCGTGGAACA TTAGCATGGA 1136
 Lys Val Gln Asp Ile Asp 315
 TGTCCCTAGAT GTTTGGAAAC TTCTTAAAAA ATGGATGATG TCTATACATG TGTAAGACTA 1196
 CTAAGAGACA TGGCCACCG TGTATGAAAC TCACAGCCCT CTCTCTTGAG CCTGTACAGG 1256
 TTGTGTATAT GTAAAGTCCA TAGGTGATGT TAGATTTCATG GTGATTACAC AACGGTTTAA 1316

FIG.1E

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CAATTGTGTA ATGATTTCCT AGAATTGAAC CAGATTGGGA GAGGTATTCC GATGCTTATG	1376
AAAAAATTAC ACGTGAGCTA TGGAAAGGGG TCACAGTCTC TGGGTCTAAC CCCTGGACAT	1436
GTGCCACTGA GAACCTTGAA ATTAAGAGGA TGCCATGTCA TTGCAAAGAA ATGATAGTGT	1496
GAAGGGTTAA GTTCTTTTGA ATTGTTACAT TGGGCTGGGA CCTGCAAATA AGTTCTTTTT	1556
TTCTAATGAG GAGAGAAAAA TATATGTATT TTTATATAAT GTCATAAGTT ATATTTCAGG	1616
TGTAATGTTT TCTGTGCAAA GTTTTGTAAA TTATATTGT GCTATAGTAT TTGATTCAAA	1676
ATATTAAAA ATGCTCACT GTGACATAT TTAATGTTTT AAATGTACAG ATGTATTAA	1736
CTGGTGCACT TTGTAATTCC CCTGAAGGTA CTCGTAGCTA AGGGGGCAGA ATACTGTTC	1796
TGGTGACCAC ATGTAGTTA TTTCTTTATT CTTTTTAACT TAATAGAGTC TTCAGACTTG	1856

FIG. 1F

TCAAAACTAT GCAAGCAAAA TAAATAAATA AAAATAAAAT GAATACCTTG AATAATAAGT 1916
AGGATGTTGG TCACCAGGTG CCTTTCAAAT TTAGAAGCTA ATTGACTTTA GGAGCTGACA 1976
TAGCCAAAAA GGATACATAA TAGGCTACTG AAATCTGTCA GGAGTATTTA TGCAATTATT 2036
GAACAGGTGT CTTTTTTTAC AAGAGCTACA AATTGTAAAT TTTGTTTCTT TTTTTTCCCA 2096
TAGAAAAATGT ACTATAGTTT ATCAGCCAAA AAACAATCCA CTTTTTAATT TAGTGAAAGT 2156
TATTTTATTA TACTGTACAA TAAAAGCATT GTCTCTGAAT GTTAATTTTT TGGTACAAAA 2216
AATAAATTG TACGAAAACC TGAAAAAAA AAAAAAAA CCGCCGCTCT 2276
AGAGGGCCCT ATTCTATAG 2295

FIG.1G

8 / 3 0

FIG.2A

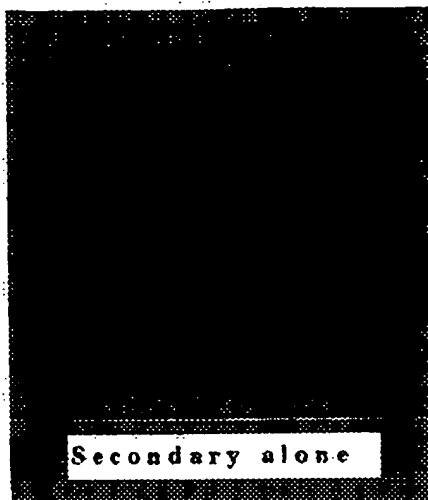


FIG.2B

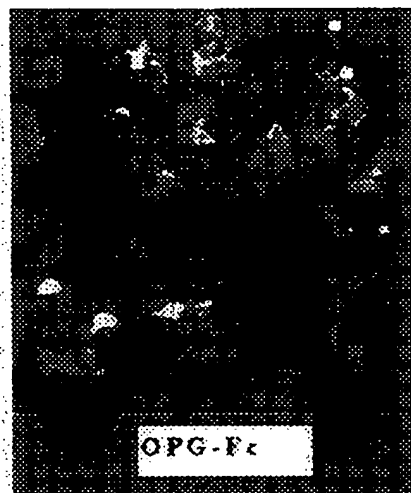
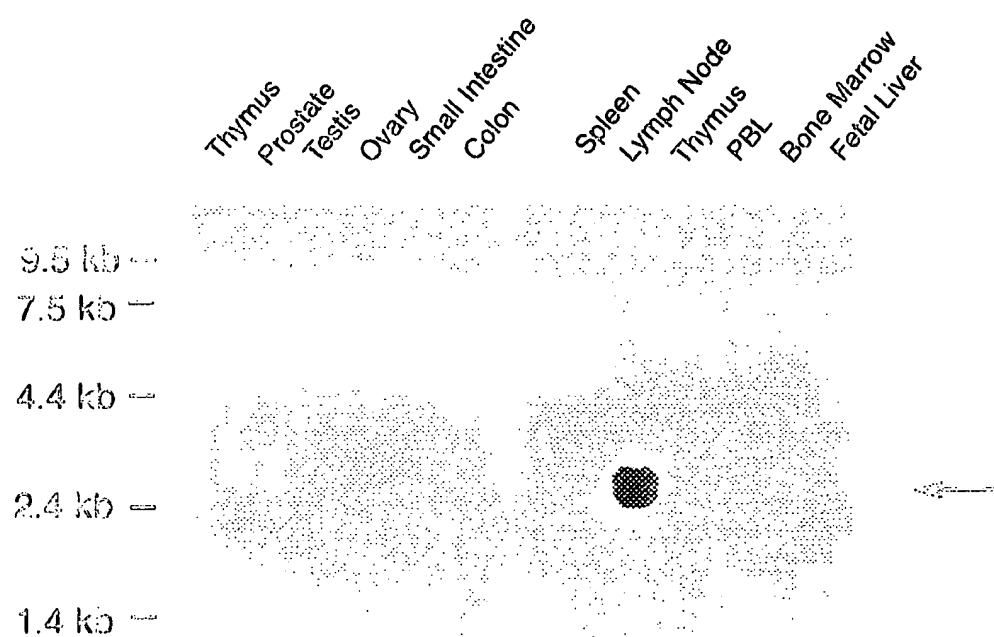


FIG.2C



9 / 3 0



10 / 30

10 30 50
AAGCTTGGTACCGAGCTCGGATCCACTACTCGACCCACGCGTCCGGCGCCCCAGGAGCC
70 90 110
AAAGCCGGCTCCCAAGTCGGCGCCCCACGTCGAGGCTCCGCCCGCAGCCTCCGGAGTTGGC
130 150 170
CGCAGACAAGAAGGGAGGGAGCGGGAGAGGGAGAGAGCTCCGAAGCAGAGGCCGAG
190 210 230
CGCCATGCGCGCGCCAGCAGAGACTACACCAAGTACCTGCGTGGCTCGGAGGAGATGGG
M R R A S R D Y T K Y L R G S E M G
250 270 290
CGGCGGCCCCGGAGCCCCGACGAGGGCCCCCTGCACGCCGCCGCGCTGCGCCGCA
G G P G A P H E G P L H A P P P A P H
310 330 350
CCAGCCCCCGCGCTCCCGCTCCATGTTTCGTGGCCCTCCTGGGGCTGGGGCTGGGCCA
Q P P A A S R S M F V A L L G L G L G Q
370 390 410
GGTTGTCTGCAGCGTCCCGCTGTTCTTCTATTTCAGAGCGCAGATGGATCCTAATAGAAT
V V C S V A L F F Y F R A Q M D P N R I

FIG.4A

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430 450 470
ATCAGAAGATGGCACTCACTGCATTATAGAAATTTTGAGACTCCATGAAATGCAGATTT
S E D G T H C I Y R I L R L H E N A D F

490 510 530
TCAAGACACAACCTCTGGAGAGTCAAGATACAAAATTAATACCTGATTCATGTAGGAGAAT
Q D T T L E S Q D T K L I P D S C R I

550 570 590
TAAACAGGCCTTTCAAGGAGCTGTGCAAAAGGAATTACAACATATCGTTGGATCACAGCA
K Q A F Q G A V Q K E L Q H I V G S Q H

610 630 650
CATCAGAGCAGAGAAAGCGATGGTGGATGGCTCATGTTAGATCTGGCCAAAGAGGAGCAA
I R A E K A M V D G S W L D L A K R S K

670 690 710
GCTTGAAGCTCAGCCTTTTGCTCATCTCACTATTAAATGCCACCGACATCCCATCTGGTTC
L E A Q P F A H L T I N A T D I P S G S

730 750 770
CCATAAAGTGAGTCTGTCCCTCTTGGTACCATGATCGGGGTTGGGCCAAGATCTCCAACAT
H K V S L S S W Y H D R G W A K I S N M

FIG.4B

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790 810 830
GACTTTAGCAATGGAAACTAATAGTTAATCAGGATGGCTTTTATTACCTGTATGCCAA
T F S N G K L I V N Q D G F Y Y L Y A N

850 870 890
CATTTGCTTTCGACATCATGAAACTTCAGGAGACCTAGCTACAGAGTATCTTCAACTAAT
I C F R H H E T S G D L A T E Y L Q L M

910 930 950
GGTGACGTCACTAAACCAGCATCAAAATCCCAAGTTCTCATACCTGATGAAGGAGG
V Y V T K T S I K I P S S H T L M K G G

970 990 1010
AAGCACCAAGTATTGGTCAGGGAATTCTGAAATTCCTATTTTATTCATAAACGTTGGTGG
S T K Y W S G N S E F H F Y S I N V G G

1030 1050 1070
ATTTTTTAAGTTACGGTCTGGAGAGGAAATCAGCATCGAGGTCTCCAAACCCCTCCTTACT
F F K L R S G E E I S I E V S N P S L L

1090 1110 1130
GGATCCGGATCAGGATGCAACATACTTTGGGCTTTTAAAGTTCGAGATATAGATTGAGC
D P D Q D A T Y F G A F K V R D I D

FIG.4C

1150 1170 1190
CCCAGTTTGTGGAGTGTTATGTATTTCTCGGATGTTGGAAACATTTTAAACAAGCC
1210 1230 1250
AAGAAAGATGTATATAGGTGTGTGAGACTACTAAGAGGCATGGCCCCCAACGGTACACGAC
1270 1290 1310
TCAGTATCCATGCTCTTGACCTTGTAGAGAAACACGCCGTATTTACAGCCAGTGGGAGATGT
1330 1350 1370
TAGACTCATGGTGTGTACACAAATGGTTTTTTAAATTTTGTAATGAATTCCTAGAAATTAAA
1390 1410 1430
CCAGATTGGAGCAATTACGGGTTGACCTTATGAGAAACTGCATGTGGGCTATGGGAGGGG

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FIG.4D

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1450 1470 1490
TTGGTCCCTGGTCATGTGCCCCCTTCGCAGCTGAAGTGGAGAGGGTGTCACTAGCGCAAT

1510 1530 1550
TGAAGGATCATCTGAAGGGGCAAAATCTTTTGAATTGTTACATCATGCTGGAACCTGCAA

1570 1590 1610
AAAAATACTTTTCTAATGAGGAGAGAAAAATATATGTATTTTATATAATCTAAAGTTA

1630 1650 1670
TATTTCAGATGTAATGTTTCTTTGCAAGTATTGTAAATTATTTGTGCTATAGTATT

1690 1710 1730
TGATTCAAAATATTTAAATGTCTGTGCTGTGACATATTTAATGTTTAAATGTACAGA

1750 1770 1790
CATATTTAACTGGTGCACTTTGTAAATTCCTGGGAAACTTGCAGCTAAGGAGGGGAA

1810 1830 1850
AAAAATGTTGTTCCCTAATATCAAAATGCAGTATATTCTTCGTTCTTTTAAAGTTAATAG

FIG.4E

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1870 1890 1910
ATTTTTCAGACTTGTCAAGCCTGTGCAAAAAATTAAATGGATGCCTTGAAATAAAG

1930 1950 1970
CAGGATGTTGGCCACCAGGTGCCCTTTTCAAAATTTAGAACTAATTGACTTTAGAAAGCTGA

1990 2010 2030
CATTGCCAAAAGGATACATAATGGGCCACTGAAATCTGTCAAGAGTAGTTATATAATTG

2050 2070 2090
TTGAAACAGGTGTTTTTCCACAAGTGCCGCAAAATTGTACCTTTTTTTTTTTCAAAATAG

2110 2130 2150
AAAAGTTATTAGTGGTTTATCAGCAAAAAAGTCCAATTTAATTAGTAAATGTTATCTT

2170 2190 2210
ATACTGTACAAATAAAACATTGCCCTTTTGAATGTTAATTTTTTGGTACAAAAATAAATTA

2230 2250 2270
TATGAAAAAATAAAAGGCGCGCTCTAGAGGGCCCTATTCCTATAG

FIG. 4F

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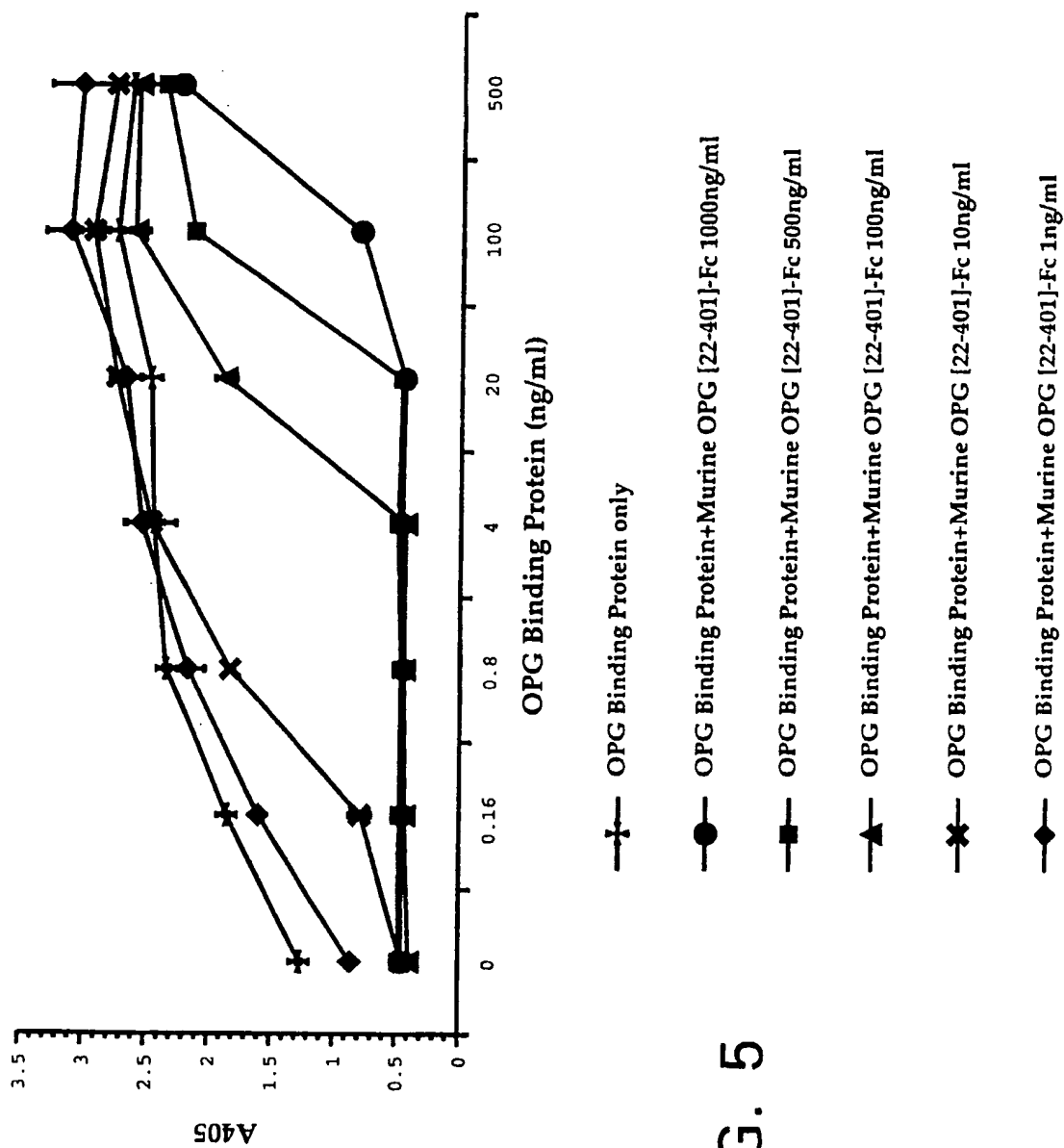


FIG. 5

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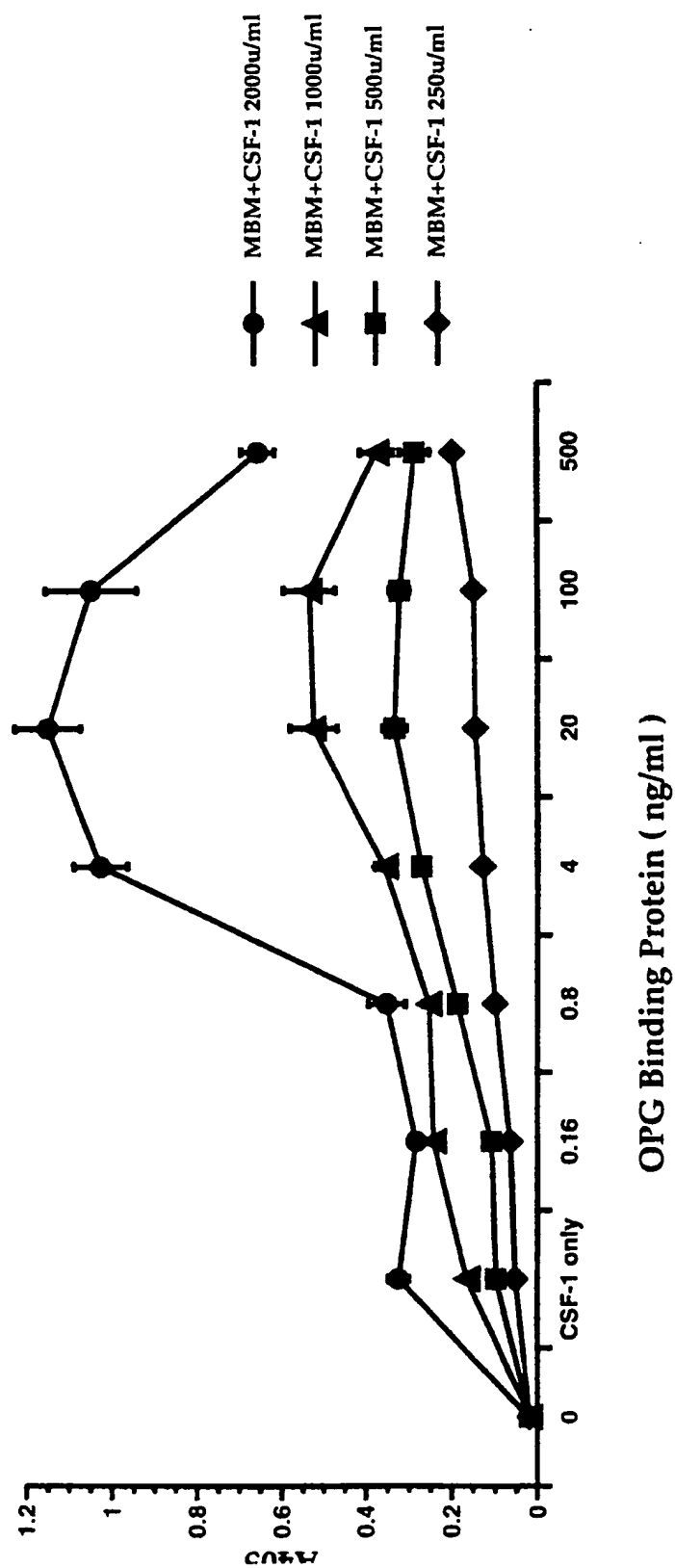
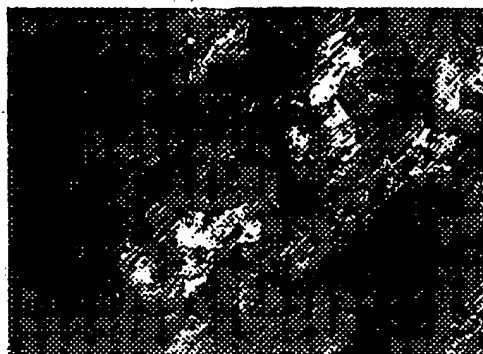
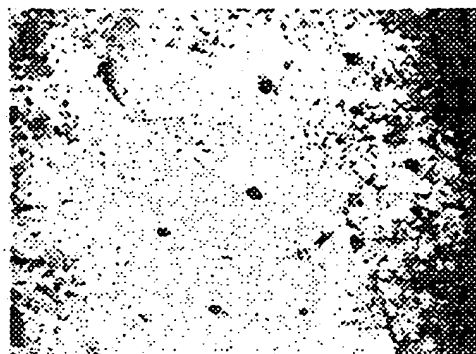
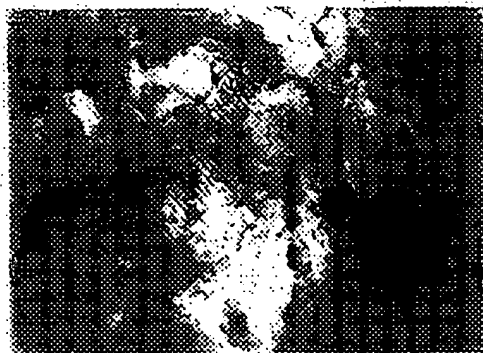
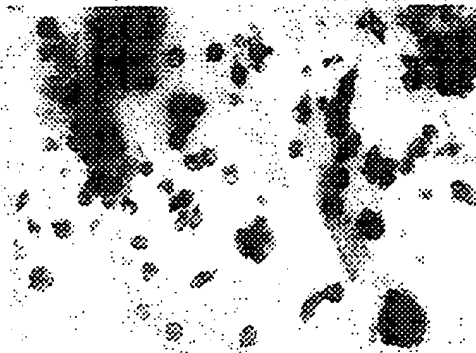
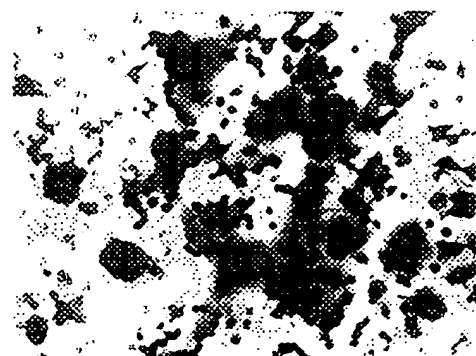


FIG. 6

FIG. 7A**Toluidine Blue Staining****TRAP staining****Bone Marrow Cells + M-CSF-1****FIG. 7B****Toluidine Blue Staining****TRAP staining****Bone Marrow Cells + OPG Binding Protein****FIG. 7C****Toluidine Blue Staining****TRAP staining****Bone Marrow Cells + M-CSF-1 + OPG Binding Protein**
SUBSTITUTE SHEET (RULE 26)

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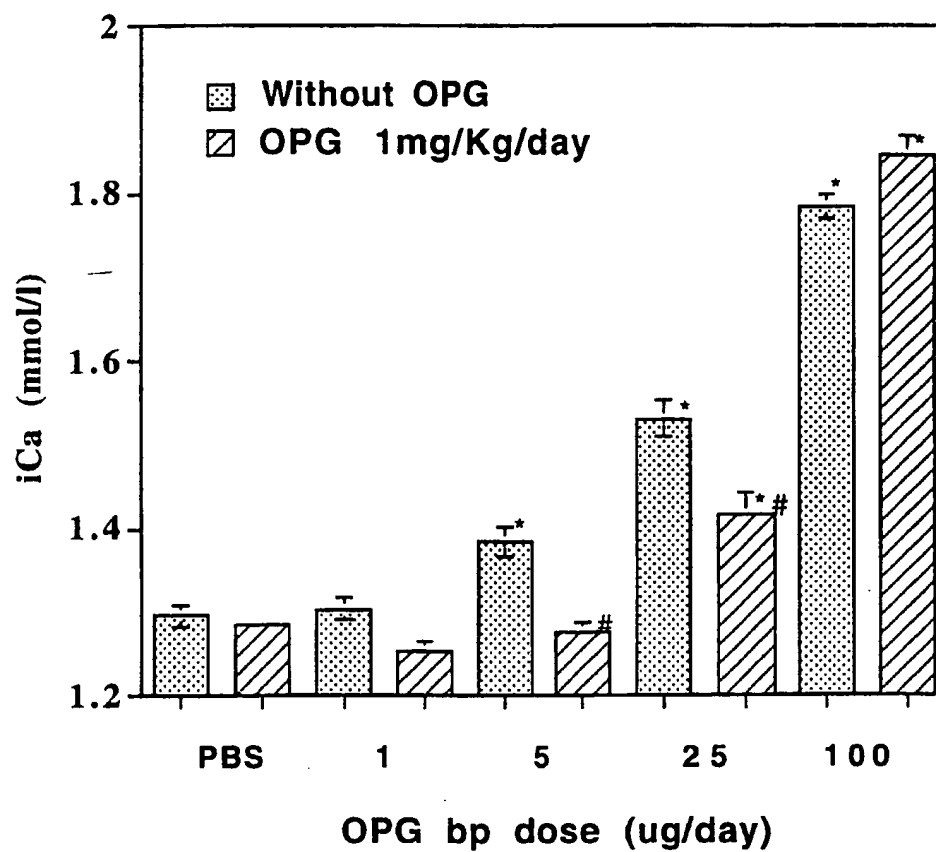


FIG. 8

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PBS

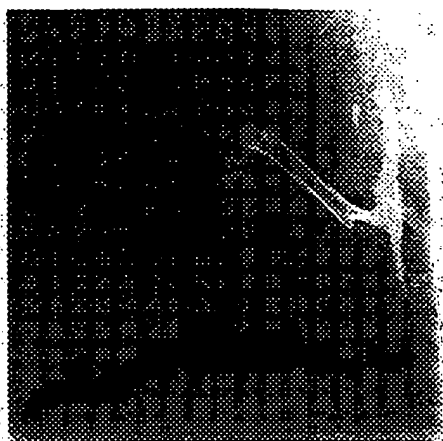


FIG. 9A

OPGbp 5ug/d

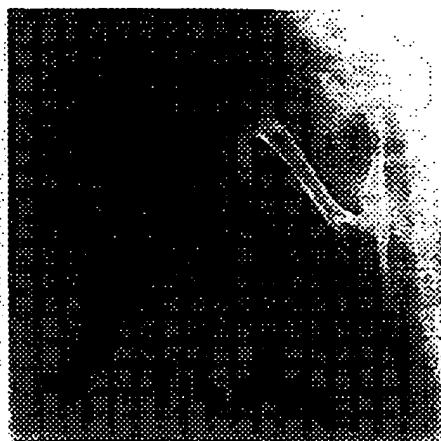


FIG. 9B

OPGbp 25ug/d

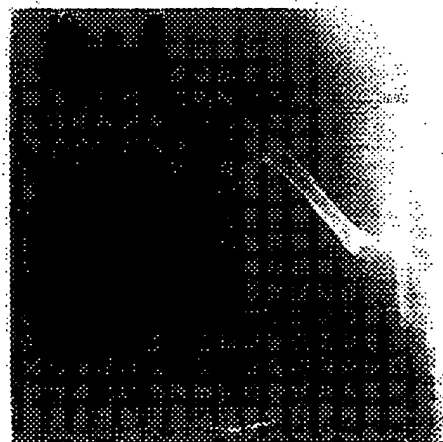


FIG. 9C

OPGbp 100ug/d



FIG. 9D

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10	30	50
ACTCGACCCACGCGTCCGCCCGCCCGCACCGGCCCATGACCCCGCGCGCGCGCGCC		
		<u>M D P R A R R R</u>
70	90	110
GCCAGCTGCCCGCGCGCTGCTGGCGCTCTGCGTGTGCTGCTCCACTGCAGGTGACTC		
<u>Q L P A P L L A L C V L L V P L Q V T L</u>		
130	150	170
TCCAGTCACTCCTCCATGCACCCAGGAGGCGCATATGAGCATCTCGGACGGTGTGCA		
Q V T P P C T Q E R H Y E H L G R C C S		
190	210	230
GCAGATGCGAACCAGGAAAGTACCTGTCTCTCTAAGTGCACTCCTACCTCCGACAGTGTGT		
R C E P G K Y L S S K C T P T S D S V C		
250	270	290
GTCTGCCCCGTGGCCCCGATGAGTACTTGGACACCTGGAATGAAGAAGATAAATGCTTGC		
L P C G P D E Y L D T W N E E D K C L L		
310	330	350
TGCATAAAGTCTGTGATGCAGGCAAGGCCCTGGTGGCGGTGGATCCTGGCAACCACACGG		
H K V C D A G K A L V A V D P G N H T A		

FIG.10A

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370 390 410
 CCCC GGT CGC GTG CT TGC ACG GCT GGT ACC ACT GGA ACT CAG ACT GCG AGT GCT GCC
 P R R C A C T A G Y H W N S D C E C C R
 430 450 470
 GCAGG AACCGG AGTGTGCACCTGGCTTCGGAGCTCAGCATCCCCTTGCAGCTCAACAAGG
 R N T E C A P G F G A Q H P L Q L N K D
 490 510 530
 ATACGGTGTGCACACCCCTGCCCTCCCTGGGCTTCTTCTCAGATGTCTTTTTCGTCCACAGACA
 T V C T P C L L G F F S D V F S S T D K
 550 570 590
 AATGCAAACCTTGGACCAACTGCACCCCTCCTTGGAAGCTAGAAAGCACACACCGGGACAA
 C K P W T N C T L L G K L E A H Q G T T
 610 630 650
 CGGAATCAGATGTGGTCTGCAGCTCTTCCATGACACTGAGGAGACCACCAAGGAGGCC
 E S D V V C S S S M T L R R P P K E A Q

FIG.10B

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670 690 710
 AGGCTTACCTGCCAGTCTCATCGTTCTGCTCCTCTTCACTCTCTGTGGTAGTAGTGGCTG
 A Y L P S L I V L L L L F I S V V V A A
 730 750 770
 CCATCATCTTCGGCGTTTACTACAGGAAGGAGGGAAGCCGTGACAGCTAATTGTGGA
 I I F G V Y Y R K G G K A L T A N L W N
 790 810 830
 ATTGGGTCAATGATGCTTGCCAGTAGTCTAAGTGGAAATAAGGAGTCCTCAGGGGACCGTT
 W V N D A C S S L S G N K E S S G D R C
 850 870 890
 GTGCTGGTTCCCACTCGGCAACCTCCAGTCAGCAAGAAAGTGTGTGAAGTATCTTACTAA
 A G S H S A T S S Q Q E V C E G I L L M
 910 930 950
 TGA CTGGGAGGAGAAGATGGTTCCAGAAAGACGGTGTGCTGGAGTCTGTGGCCCTGTGTGTG
 T R E E K M V P E D G A G V C G P V C A
 970 990 1010
 CGGCAGGTGGGCCCTGGGCAGAAAGTCAGAGATTCTAGGACGTTCACTGTGTCAGCGAGG
 A G G P W A E V R D S R T F T L V S E V

FIG.10C

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1030 1050 1070
TTGAGACGCAAGGAGACCTCTCGAGGAAGATTCCACAGAGGATGAGTACACGACCGGC
E T Q G D L S R K I P T E D E Y T D R P
1090 1110 1130
CCTCGACGCCCTTCGACTGGTTCACTGCTCCTAATCCAGAGGGAAGCAATCTATACCCC
S Q P S T G S L L L I Q Q G S K S I P P
1150 1170 1190
CATTCAGGAGCCCTGGAAGTGGGGAGAACGACAGTTTAAGCCAGTGTTCACCGGA
F Q E P L E V G E N D S L S Q C F T G T
1210 1230 1250
CTGAAAGCACGGTGGATTCTGAGGGCTGTGACTTCACTGAGCCTCCGAGCAGAACTGACT
E S T V D S E G C D F T E P P S R T D S
1270 1290 1310
CTATGCCCGTGTCCCCCTGAAAAGCACCTGACAAAAGAAATAGAAGGTGACAGTTGCCTCC
M P V S P E K H L T K E I E G D S C L P
1330 1350 1370
CCTGGGTGGTCAGCTCCAACTCAACAGATGGCTACACAGGCAGTGGGAACACTCCTGGGG
W V V S S N S T D G Y T G S G N T P G E

FIG.10D

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1390 1410 1430
AGGACCATGAACCTTTCCAGGGTCCCTGAAATGTGGACCATTGCCCCAGTGTGCCTACA
1450 1470 1490
D H E P F P G S L K C G P L P Q C A Y S
GCATGGGCTTTCCAGTGAAGCAGCAGCCAGCATGGCAGAGCGGGAGTACGGCCCCAGG
1510 1530 1550
M G F P S E A A A S M A E A G V R P Q D
ACAGGGCTGATGAGAGGGGAGCCTCAGGGTCCGGAGCTCCCCAGTGACCGCACCTG
1570 1590 1610
R A D E R G A S G S G S P S D Q P P A
CCTCTGGGAACGTGACTGGAACAGTAACTCCACGTTTCATCTCTAGCGGCAGGTGATGA
1630 1650 1670
S G N V T G N S N S T F I S S G Q V M N
ACTTCAAGGGTGACATCATCGTGGTGTATGTTCAGCCAGACCTCGCAGGAGGGCCGGGTT
1690 1710 1730
F K G D I I V V Y V S Q T S Q E G P G S
CCGCAGAGCCCGAGTCGGAGCCCGTGGGCCGCCCTGTGCAGGAGGAGACGCTGGCACACA
A E P E S E P V G R P V Q E E T L A H R

FIG.10E

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1750 1770 1790
GAGACTCCTTTGCGGGCACCGCGCGCGCTTCCCCGACGTCTGTGCCACCGGGGCTGGGC
D S F A G T A P R F P D V C A T G A G L
1810 1830 1850
TGCAGGAGGCGGACCCCGCAGAGGACGGACATCGCGCGCGGTGCAGGAGCAGG
Q E Q G A P R Q K D G T S R P V Q E Q G
1870 1890 1910
GTGGGCGCAGACTTCACTCCATACCCAGGGTCCGGACAATGTGCAGAATGACCTCACC
G A Q T S L H T Q G S G Q C A E
1930 1950 1970
TTCTCTGTGCCCCTGGGTGCAGGGCACCAAGTGCCTTTCCAAAAACATGGTGTAGCTAGC
1990 2010 2030
CACTGTGCACCTCCTCACTGGTGCAGGCTGCTGGCATGGTGATGGAGCCCACCTCTCACT
2050 2070
TCCTCCAGTGCCCCCTCTCCTCTGCCTCCTAC

FIG.10F

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FIG. 11A



FIG. 11B



FIG. 11C



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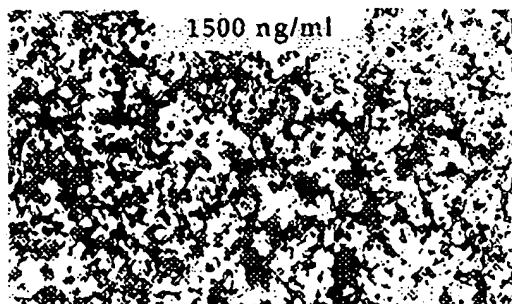


FIG.12A

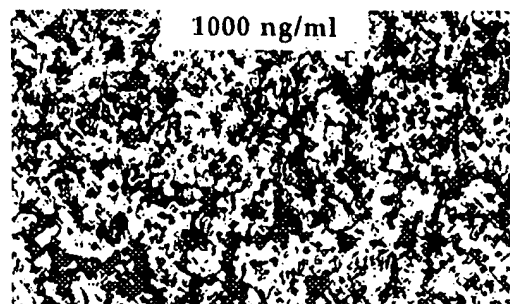


FIG.12B

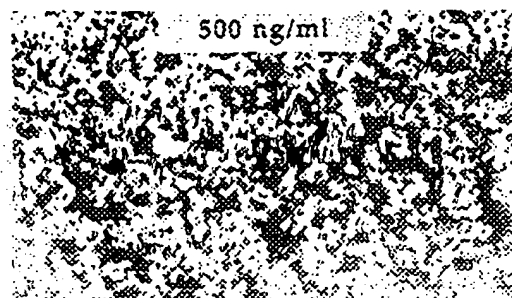


FIG.12C

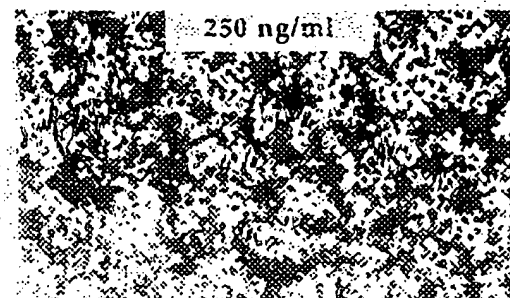


FIG.12D



FIG.12E

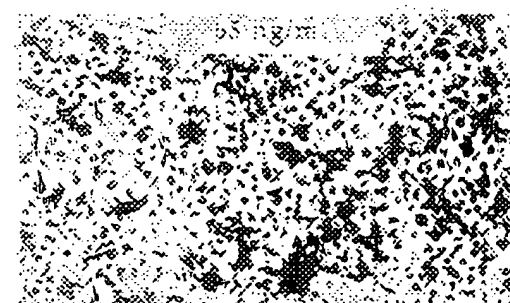


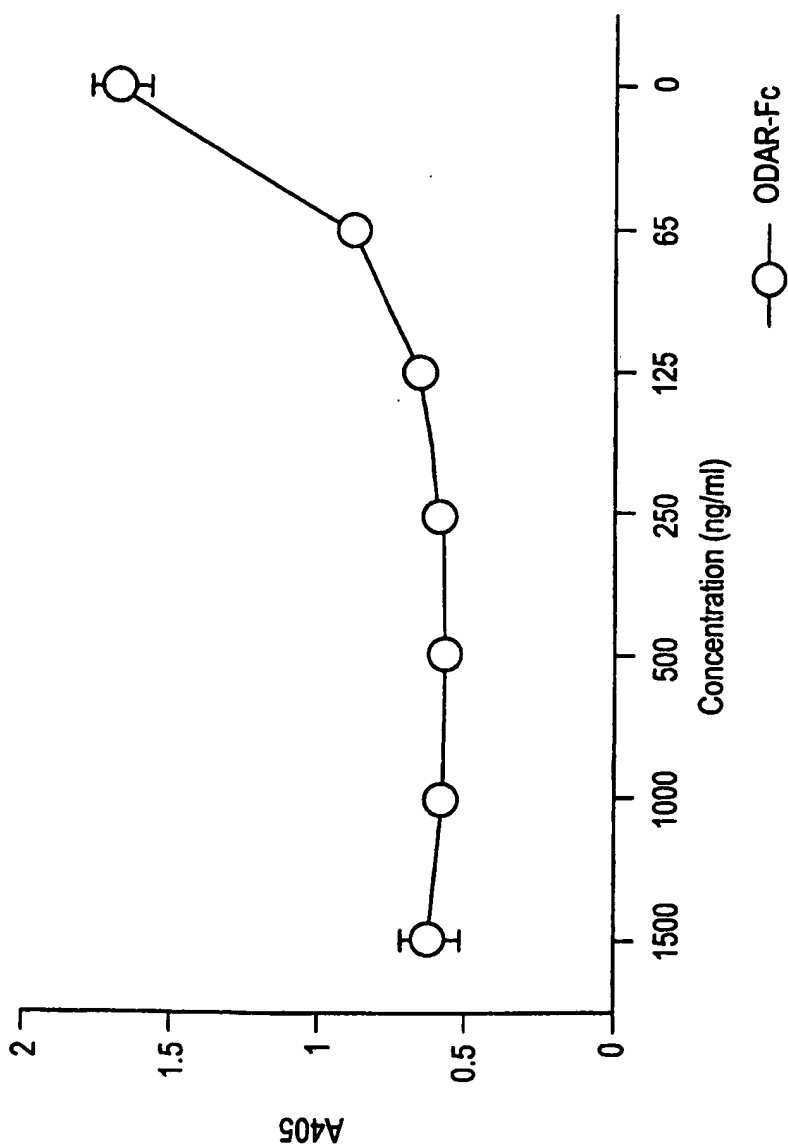
FIG.12F



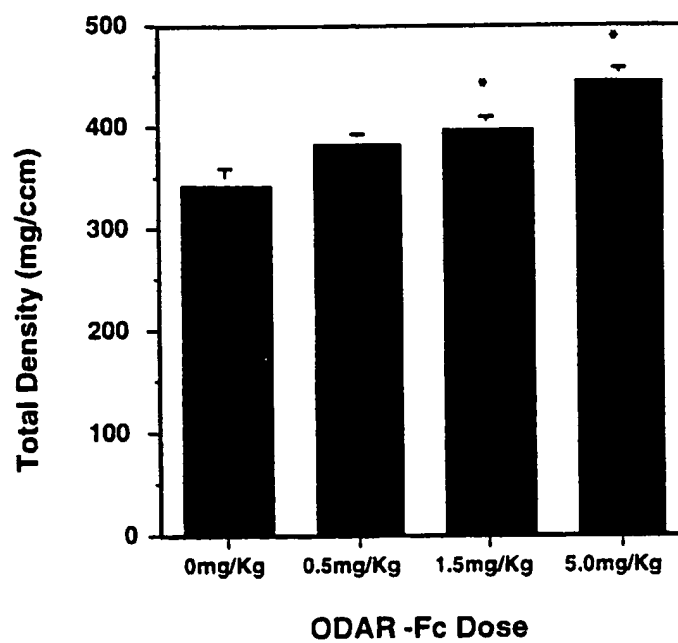
FIG.12G

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FIG. 12H



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* Different to vehicle treated control $p < 0.05$.

FIG.13

Applicant's or agent's file
reference number

A-451B

International application No. US98/07584

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Address of depositary institution (including postal code and country) 10801 University Boulevard Manassas, VA 20110-2209 U.S.A.	
Date of deposit June 13, 1997	Accession Number 98457
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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/07584

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C12N15/11 C12N1/21 C12N5/10 C07K14/705
C07K16/28 G01N33/50 G01N33/577 G01N33/68 A61K31/70
A61K38/17 A61K39/395

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K G01N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	YASUDA H ET AL: "Osteoclast differentiation factor is a ligand for osteoprotegerin/osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA., vol. 95, 31 March 1998, pages 3597-3602, XP002076248 see the whole document	1-42
E	WO 98 28426 A (IMMUNEX CORP) 2 July 1998 Seq Id Nr 11 and 13 -/--	1-27, 29-33, 37, 38, 42

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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"&" document member of the same patent family

Date of the actual completion of the international search

7 September 1998

Date of mailing of the international search report

25/09/1998

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Inter national Application No

PCT/US 98/07584

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
E	WO 98 25958 A (SCHERING CORP) 18 June 1998 See seq Id Nr 2 ---	1-27, 29-33, 37,38,42
X	DATABASE EMBEST16 E.M.B.L. Databases Accession Number: AA170348, 3 January 1997 XP002076291 see abstract ---	1
A	JIMI E ET AL: "Osteoclast function is activated by osteoblastic cells through a mechanism involving cell-to-cell contact" ENDOCRINOLOGY, vol. 137, no. 8, August 1996, pages 2187-2190, XP002076247 ---	
A	SMITH C A ET AL: "THE TNF RECEPTOR SUPERFAMILY OF CELLULAR AND VIRAL PROTEINS: ACTIVATION, COSTIMULATION, AND DEATH" CELL, vol. 76, 25 March 1994, pages 959-962, XP002029050 -----	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 98/07584

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9828426 A	02-07-1998	WO 9828424 A	02-07-1998
WO 9825958 A	18-06-1998	NONE	